

# Cholinergic System Aggravates Sik-1/Sik-3 Mediated Suppression Of Ca2+ Pathway Associated Oxidative Stress-Inflammatory Axis In Neurotoxic Alzheimer's Type Dementia

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#### Abstract:

The AD is an age-related neurodegenerative disorder characterized by progressive anterograde amnesia, cerebral atrophy, functional decline, and eventual death. among people of ages ranging from 60 to 84, 24.3 million are suffering from AD. with the number of cases estimated to rise to nearly 106 million by 2050. The pathophysiology of AD includes both structural and functional abnormalities including multiple anatomical lesions occur in the brain, including the appearance of senile plaques consisting of Aβ and neurofibrillary tangles containing phosphorylated tau, and major synaptic changes leads to substantial loss of synaptic profiles. In AD, there are also significant oxidative stress- inflammation leads to mitochondrial abnormalities and severe synaptic damage and neuronal death due to overactivation of Ca2+ pathway mediated by various factors including glutamate hyperactivity. Furthermore, the cholinergic system activation achieving the equilibrium of overactivated Ca2+ path and further activate downstream salt-inducible kinases (SIK1/SIK3) belong to AMP-activated protein kinase (AMPK) family, and functions mainly involve in regulating energy response-related physiological processes, such as gluconeogenesis and lipid metabolism. However, compared with another well-established energy-response kinase AMPK, SIKs roles in human diseases, especially in AD are rarely investigated. Thus, in this review our primary aim to investigate the Ca2+ pathway associated increase in oxidative stress- neuroinflammatory axis in AD progression and the effectiveness of cholinergic system via aggravation of SIK-1/SIK-3 mediated suppression of progression of AD is more efficient therapeutic strategy.

**Keywords:** Alzheimer's disease, Neurodegeneration, Oxidative stress- Inflammatory axis, Aβ peptides, Tau protein, Glutamate, Ca2+ pathway, Donepezil, SIKs, ROS, TNF-α, Interleukins, NF-κB signaling

#### Introduction:

Today, aging human populations around the globe are facing an epidemic of Alzheimer's disease (AD). Furthermore, among people of ages ranging from 60 to 84, 24.3 million are suffering from AD(1), with the number of cases estimated to rise to nearly 106 million by 2050 (2). The AD is an age-related neurodegenerative disorder characterized by progressive anterograde amnesia, cerebral atrophy, functional decline, and eventual death. In addition to, AD is the most common dementia type followed by vascular dementia(3). Dementia is a cognitive disability characterised by a decline in mental ability and globally occurs every 3 s. As dementia progressively leads to complete loss of autonomy requiring a permanent support, this disease represents a tremendous social and economic cost for our societies(4). Apart from genetic and socio-demographic factors such as gender and educational level, some modifiable factors such as vascular risk factors(5) or drugs consumption(6) are suspected to be associated with the risk of dementia. Clinical symptoms of AD include progressive memory decline, impaired executive function and difficulties executing routine daily activity; early symptoms of AD onset include changes in thinking or unconscious behavior, memory impairment with respect to new information, and dysfunctional changes in language and speech(7). In addition, 20 to 30% of early AD patients show significant depressive symptoms and mood changes (8). Patients in advanced stages of AD suffer from severe memory loss, hallucinations, disorientation, and lack self-sufficiency, where individuals eventually die due to respiratory syndrome(9). Although possible pharmacological interventions such as donepezil hydrochloride, a cholinergic system activator have been suggested, there are no popular therapies that decelerate or terminate its progression(10, 11). Moreover, recent study also explores blocking of cholinergic system with anti-cholinergic type of drugs increase the risk of incident mild cognitive impairment and cognitive decline, and effects were significantly enhanced among individuals with genetic risk factors and CSF-based AD pathophysiological markers(12). Thus, conforming the involvement of cholinergic system in AD. However, the treatment has limited efficacy, are symptomatic, and are unable to decelerate disease progression (13, 14) possibly because they are administered at advanced AD stages when synapse loss is too pronounced. This lack of disease modifying treatment is due to poor identification of effective biomarkers for early diagnosis. Accordingly, intense efforts are ongoing to understand neurological changes associated with AD progression. Thus, in this review our primary aim to investigate the Ca2+ pathway associated increase in oxidative stress- neuroinflammatory axis in AD progression and the effectiveness of cholinergic system via aggravation of salt inducible kinas-3 (SIK-3) mediated suppression of progression of AD is more efficient therapeutic strategy.

#### Selection of literature for review:

The potentially relevant studies were retrieved from the ScienceDirect/Medline/PubMed/Public library of science/Mendeley/Springer link and Google Scholar. Multiple keywords were used for the literature search both alone as well as in combination. Some of the important keywords used for literature search were 'Definition of AD', 'Epidemiology of AD', 'AD associated inflammation', 'Mechanism of apoptosis in AD', 'Involvement of Aβ in neurodegeneration, 'AD associated glutamate receptor activation', 'Risk factor for AD', 'Pathogenesis of AD', 'Reactive oxygen species mediated mitochondrial defect, 'Role of donepezil for improving inflammation', 'Mechanism of action of donepezil', 'salt-inducible kinases association in inflammatory signalling', 'Cellular metabolism in AD', 'Inflammasome pathway in cell death', 'Cholinergic system and calcium pathway', 'Innate and Adaptive immunity in AD'. Only articles with English language were considered in the present study. The reference lists of retrieved articles were also screened to find relevant articles that were not identified by the initial search strategy.

#### Pathophysiology of Alzheimer's disease:

The pathophysiology of AD includes both structural and functional abnormalities. As AD progresses, multiple anatomical lesions occur in the brain, including the appearance of senile plaques consisting of Aß and neurofibrillary tangles containing phosphorylated tau, and major synaptic changesleads to substantial loss of synaptic profiles(15). In AD, there are also significant oxidative stressinflammationleads to mitochondrial abnormalities and severe synaptic damage and neuronal death(15). Glutamate is the most abundant excitatory neurotransmitter in the mammalian Central Nervous System (CNS), acting at ionotropic and metabotropic glutamate receptors. Ionotropic glutamate receptors (iGluRs), responsible for fast neuronal communication at excitatory synapses, comprise three subfamilies:  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxasolepropionic acid (AMPA) receptors, kainate receptors, and NMDARs(16). NMDARs are glutamate-triggered ion-gated cationic channels which play a pivotal role in excitatory synaptic transmission, plasticity and excitotoxicity in the nervous system(17). Seven NMDAR subunits have been characterized in total for GluN1, GluN2 and GluN3 subtypes: GluN1, GluN2A through D, and GluN3A and B. Structurally, functional NMDAR comprises two GluN1 and GluN2 or GluN3 subunits which can form a  $Ca^{2+}$ -permeable ion channel(18)It is extensively distributed in the CNS whereas it is almost exclusively located intracellularly. Activation of NMDARs through the accumulation of AB likely occurs during early stages of disease progression(19). Similar to NMDA stimulation, AB evoke immediate cellular  $Ca^{2+}$  influx through the activation of GluN2B-containing NMDARs in primary neurons. Glutamate is prevalent in neocortical and hippocampal pyramidal neurons and plays a role in synaptic plasticity, learning, and memory consolidation. Synaptic glutamate signaling is tightly regulated by clearance through high-affinity excitatory amino acid transporters(20). However, postmortem analysis shows that vesicular glutamate transporter 1 boutons were elevated in pre-clinical AD cases(21) while glutamate transporters were decreased in AD patients(22). Indicating mechanisms responsible for glutamatergic regulation are altered during different stages of disease progression. A contributing factor to this dysregulation is the accumulation of soluble A $\beta$  isoforms that initiates synaptic dysfunction causing the eventual neurodegeneration(23, 24). Moreover, the underlying cause of selective hippocampal CA1 neuronal degeneration is a result of glutamate-mediated excitotoxic mechanisms involving excessive calcium influx through NMDAR activation, mitochondrial dysfunction, and reactive oxygen species. These events culminate in necrotic cell loss that releases more glutamate into the extracellular space thus propagating damage to surrounding neurons(25). In addition to that, tau phosphorylation is regulated by multiple protein kinases and phosphatases. Tau kinases can be classified to two categories: 1) Ser/Thr kinases such as CDK5, glycogen synthase kinase  $3\beta$  (GSK3 $\beta$ ), mitogen-activated protein kinase,  $Ca^{2+}/calmodulin-dependent$  protein kinase II, microtubule-affinity regulating kinase, protein kinase A (PKA), protein kinase C, Akt, TTBK1/2, CK1, DYRK1A, and 2) tyrosine kinases including Fyn, Src, Syk and c-Abl(26). Tau is dephosphorylated by protein phosphatase 1 (PP1), PP2A, PP2B and PP5(27-29). Tau hyperphosphorylation may result from imbalanced activity or expression of tau kinases and protein phosphatases. In support of this, increased GSK3ß expression and CDK5 activity, decreased expression of PP1 and PP2A, and decreased PP2A activity has been observed in

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specific brain regions in AD patients(30-34). Since hyperphosphorylated tau species are enriched in NFTs, strategies to suppress tau phosphorylation may be a viable therapeutic strategy in AD and other related tauopathies. Unfortunately, attempts to target hyperphosphorylated tau or inhibit tau kinases have not seen success so far. Therefore, understanding changes to glutamatergic signaling at different stages of AD progression could lead to the development of disease-stage specific therapeutics (Figure 1).



Figure 1: The mechanistic bio-molecular involvement of calcium pathway mediated aggravation of oxidative stress inflammatory axis defect and Alzhiemer's disease progression

## Involvement of ca2+ pathway associated oxidative stress- inflammatory axis in disease progression:

In the biomolecular pathology of AD accompanied by neuroinflammatory reactions, oxidative stress, and free radical formation probably associated with mitochondrial dysfunction, excitotoxic reactions, alterations in cholesterol metabolism and lipid rafts, deficiencies in neurotransmitters (especially acetylcholine) and neurotrophic factor function, defective activity of the ubiquitin-proteasome, and chaperone systems and cerebrovascular dysregulation(35). Accumulating evidence demonstrates that the toxicity is principally mediated by excessive Ca<sup>2+</sup> entry, primarily through NMDARs(36-39). since NMDARs have a much higher permeability for calcium ions compared to other iGluRs(40). In this regard,

the modest depolarization of the postsynaptic membrane and other factors that relieve the Mg<sup>2+</sup> blockade can activate NMDARs in a mild and chronic way, which causes the prolonged Ca<sup>2+</sup> influx into the postsynaptic neuron. The pathological level of Ca<sup>2+</sup> signaling leads to gradual loss of synaptic function and ultimate neuronal cell death, which correlates clinically with the progressive decline in cognition/memory and the development of pathological neural anatomy seen in AD patients, and this, in turn, rationalizes the clinical trial of memantine, an NMDAR antagonist, as a symptomatological and neuroprotective treatment for AD(41-43). Furthermore, a pathological increase in the amount of AB explore the functional and morphological changes in glial cells, including calcium dysregulation. In fact, microglia and astrocytes are activated close to senile plaques to internalize and break down A $\beta$  (44). This cellular activation may result in an inflammatory response and OS, playing a dual role in the pathophysiology of AD with both detrimental and neuroprotective results. Inflammatory mediators (i.e., bradykinin) may increase intracellular calcium concentration via nicotinic receptors and PI3K-Akt pathway in cultured astrocytes(45). In the CNS, nAChRs are expressed in neurons and glial cells, including microglia, oligodendrocytes and astrocytes, with highest expression in astrocytes among the glial cells that release Ca2+ dependent release of glutamate neurotransmitters(46, 47). Oxidative stress manifests early in AD, which supports the notion that oxidative stress may drive AB-induced AD pathogenesis (48). AB peptides can induce ROS production from mitochondria, leading to release of cytochrome c and apoptosis-inducing factor, thereby driving mitochondrial dysfunction, cell apoptosis and neuronal loss (49). However, AB interacts with several types of surface receptors in astrocytes which leads to calcium entry through the activation of calcium channels, including purinergic receptor P2Y1(50)and glutamate metabotropic receptor mGluR5(51, 52). Furthermore, Aβ can induce abnormal elevations in extrasynaptic glutamate levels and subsequent extrasynaptic N-methyl-D-aspartic acid receptor (NMDAR)-mediated excitotoxicity. In addition to excreted glutamate can activate extrasynaptic NMDAR in neurons residing within neuron/astrocyte conjugates, resulting in Ca<sup>2+</sup> efflux. This triggers multiple events, including mitochondrial dysfunction, caspase 3 activation, tau hyperphosphorylation, and excessive production of NO, ROS and VEG-F. These events result in damage to dendritic spines and neuronal synapses, disrupting neuronal/astrocytic communication(53, 54). Additionally, oligomeric AB can disrupt intracellular calcium balance, impair mitochondria dysfunction, and induce the production of reactive oxygen species (ROS). Mitochondria are the primary source of intracellular ROS and excessive ROS accumulation can further aggravate oxidative stress and mitochondrial DNA damage and dysfunction. All of these events eventually lead to neuronal apoptosis and cell death(55). As several study reports that psychological and chronic stress long-term activation of the HPA axis eventually leading to the secretion of glucocorticoid into the bloodstream, where blood glucocorticoid enters the brain through the BBB to activate the glucocorticoid receptor (GR in human) and mineral corticosteroid receptor (MR in mice) accompanied by permanent depletion of receptors and loss of hippocampal neurons suggesting the contributing factor in the pathogenesis of AD and other neurodegenerative diseases(56-59). Oxidative stress is further exacerbated by glutamate excitotoxicity through upregulation of glutamate receptors. In response to this, neurons and glial cells (i.e., microglia, astrocytes, and oligodendrocytes) mediate neuroimmune reactions through interactions with neuroimmune factors such as Toll-like receptors (TLRs), high-mobility group protein box 1 (HMGB1), and pro-inflammatory cytokines. In addition, A $\beta$ -induced inflammation may also contribute to tau pathology. Aβ plays a primary role in activating several innate immune pathways, causing inflammatory response

and releasing inflammatory cytokines, such as interleukin-1 $\beta$  (IL-1 $\beta$ )(60, 61). Microglia may express other putative A $\beta$  receptors, including Toll-like receptor 2/4 (TLR2/4) (62), complement receptor 3 (CR3)(63), Fc y receptors IIb (FcyRIIb)(64), CD36(65, 66), advanced glycation end product receptor (RAGE)(67). These receptors cooperatively bind, internalize and clear A $\beta$ , in addition to modulating microglial activation mediated neurotoxicity and memory impairment triggered by AB.The TLR can participate in AD pathogenesis and induce microglial inflammation phagocytosis through interactions with A $\beta$  (68). Specifically, TLR4 activation induced NF- $\kappa$ B nuclear translocation, leading to the production of proinflammatory mediators(69). Moreover, activated microglia can also release proinflammatory cytokines including IL-1 $\beta$ , IL-6, as well as tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) in AD, and enhance oxidative stress through induced ROS generation(70, 71). It has been shown that IL-1 $\beta$ , lipopolysaccharide (LPS), prostaglandin E2 and tert-butyl hydroperoxide can reduce the microglial phagocytosis, thereby enhancing A $\beta$  aggregation(72). The activated astrocytes in AD can aggravate inflammation by producing proinflammatory cytokines and active nitrogen and oxygen species (RNS, ROS) which interfere with synaptic germination and axonal growth(73, 74). Additionally,  $A\beta$  can indirectly induce glutamatergic toxicity by reducing distribution of the astrocytic glutamate transporter, GLT1 (EAAT2, SLC1A2) to the cell surface(75). Moreover, pathogenic tau species can also activate microglia and astrocytes independently of AB. Tau-dependent microglial activation can further enhance secretion of proinflammatory cytokines such as IL-1 $\beta$ , IL-6, and TNF- $\alpha$ (76-78), along with recent transcriptomic studies exhibiting the role of NF-κB activation and NLRP3-ASC for the inflammatory process in AD. Notably, Aβ also can activate NF-κB signaling and the NLRP3-ASC inflammasome (79, 80), suggesting that Aβ and tau share common mechanisms in microglia activation along with inflammatory events in AD. Furthermore, tau can be ubiquitinated by several E3 ligases including TNF receptor-associated factor 6 (TRAF6), induces Lys63-linked tau polyubiquitination and 26S proteasome-mediated tau degradation. Thus, consequence of ubiquitination on tau degradation and pathogenesis exert important relationship with TRAF6 activity. Henceforth, the achieving equilibrium in overall Ca<sup>2+</sup> level in CNS, mediated by NMDAR leading to damage-associated learning and long-term memory deficits may be disease-stage specific therapeutics and help in prevention of overall progression of disease (Figure 2).



Figure 2: Explore cholinergic system aggravates SIK-1/SIK-3 mediated suppression of oxidative stressinflammatory axis in neurotoxic Alzheimer's type dementia

## Relationship of salt-inducible kinases (SKIs) with factors contributing alzheimer's disease:

Salt-inducible kinases (SIKs) belong to AMP-activated protein kinase (AMPK) family, and functions mainly involve in regulating energy response-related physiological processes, such as gluconeogenesis and lipid metabolism. However, compared with another well-established energy-response kinase AMPK, SIK roles in human diseases, especially in AD are rarely investigated.

Salt-inducible kinase (SIK) was first identified in the adrenal glands of high salt diet-fed rats in 1999(81). Further, the SIK family members, including SIK1–SIK3, are characterized as serine/threonine kinases that belong to AMP-activated protein kinase (AMPK) family(82, 83). Later, SIKs have shown self-phosphorylation, and play an important role in regulating adrenocortical function under the stimulation of high salt or adreno-cortico-tropic-hormone (ACTH)(81).SIK1 gene is located in human chromosome 21, while SIK2 and SIK3 genes are both located on chromosome 11(82). Beyond that, SIKs are also composed of a central sucrose non fermenting (SNF-1) homology (SNH) domain, and a long C-terminal domain (84, 85). The N-terminal KD contains a LKB1 phosphorylation site and is relatively conserved across SIK family. However, the SNH domain is distinct in SIKs, specifically, the similarity percentage of SIK2 and SIK3 compared that of SIK1 is 70% and 37% respectively. The C-terminal domain contains multiple protein kinase A (PKA) phosphorylation sites and is highly conserved between SIK1 and SIK2 (85). Of note, the SIK1 is abundantly expressed in the adrenal cortex, as well as in the adipose and neural tissues(83, 86, 87), while both SIK2 and SIK3 are ubiquitous in humans and mainly expressed in adipose and neural tissues, respectively(83).

In recent years, although the roles of SIKs in AD have drawn much attention due to their association

with neuroinflammation and undergo apoptosis through NF-kB signaling pathway.

It has been seen that alcohol consumption not only causes neuroinflammation and increase a variety of immune-related signaling processes but also responsible for Alzheimer's type dementia. Furthermore, A study establishes not only that SIK1 depletion promoted alcohol-induced neuroinflammation and microglial apoptosis, but also that the NF-κB signaling pathway is required for its activity(88). There are indications that overexpression of SIK-3 and SIK-1 but not SIK-2 inhibits NF-kB activation in response to TLR4 stimulation and decreases production of proinflammatory cytokines(89). Furthermore, SIK-3, but not SIK-1 and SIK-2, deficiency is responsible for the profound inflammatory response on lipopolysaccharide(90). In addition to, SIK1and SIK3 has been reported to interacts with TGF-β-activated kinase 1-binding protein 2 (TAB2) and interrupt the complex of TAB2-TNF receptor-associated factor 6 (TRAF6)(91). Thereby inhibits TLR4-mediated NF-κB signaling which is crucial for ubiquitination on tau in AD(91). Although SIK-1 and SIK-3 activation seems to be anti-inflammatory, the activation of SIK-2, on the contrary, is harmful and suppress neuronal survival both in cell culture and in whole animals after ischemic stroke(92). Meanwhile, CRTC and class IIa HDACs, two important downstream substrates of SIKs, negatively regulate NF-kb signaling pathway. However, a study exploring the effectiveness of bosutinib in the model of chronic inflammation in AD via SIK-2 inhibition mediated dephosphorization of CRTC-3, inducing the translocation of CRTC-3 to the nucleus. The translocation activates CREB, promoting CREB-dependent gene transcription and inhibiting NF-κB(93).

It is well established that insulin-like growth factor I (IGF-I) enhances SIK1 activity in a PI 3-kinasedependent manner via PDK1. further evidence suggested WNK1 mediated activation of SIK1 by IGF-I(94) where the later one is essential for progression of angiogenesis and neurogenesis during neurodegeneration(95). SIK1 can be activated by an increase of cytosolic  $Ca^{2+}$  activity, an effect presumably mediated by calmodulin-dependent protein kinase kinase (CaMKK)(96). SIKs are considered rapid turnover proteins due to the phosphorylation by PKA, PKC, and tyrosine kinase in their C-terminal region(97). SIKs could be specifically activated by the sodium homeostasis(82). As a result, sodium intake-induced calcium influx affected by Na<sup>+</sup>/Ca<sup>2+</sup> exchange system (NCE1), could cause CaMKmediated SIK1 phosphorylation and activation(96, 98, 99). Phospholipase C (PLC) can boost Ca<sup>2+</sup> influx from endoplasmic reticulum (ER) to the cytoplasm via inositol triphosphate (IP3) receptor, thus activating the CaMK, which leads to the phosphorylation and activation of SIK2 at Ser358(100). Interestingly, a study found that CaMK I/IV phosphorylated SIK2 at Thr484, leading to SIK2 degradation and promoting CREB-mediated transcription(101). SIK-1 directly phosphorylate some specific substrates, including CRTC/CREB and PPase methylesterase-1 (PME-1) to involve in metabolic homeostasis(99). SIK1 was first found to inhibit gluconeogenesis in the hepatocytes, and its mRNA and protein levels under fasting conditions increased fourfold relative to feeding conditions(102). In-addition to, SIK3 plays a key role in regulating gluconeogenesis and expression of GLUT1 (provide glucose to damage neurons) rather than SIK2 the process could be helpful during neurodegenerative process of AD(103, 104). Moreover, overexpression of SIK1 result repression of lipogenic genes (FAS, ACC2 and SCD1) &SIK3 has been reported as a new energy regulator by promoting lipid storage in Drosophila through compromising the activity of HDAC4 and CRTC(97, 105).In addition, SIK3 also regulated cholesterol and bile acid metabolism by combining with retinoic acid metabolism and might alter energy storage in mice(106). Meantime, SIK2 also promotes cholesterol synthesis by upregulating SREBP2 expression, to transcriptionally elevate cholesterol synthetase, 3-hydroxy-3-methyl-glutaryl-coenzyme A reductase

(HMGCR)(107). Henceforth, the process of lipid metabolism, mediated by SIK1/SIK3 may helpful in utilization/suppression of formation of A $\beta$  and taueffective in prevention of progression of AD.

#### Potential of cholinergic drugs and SIK--1/SIK-3 mediated suppression of Alzheimer's disease

Previous studies have shown that cognitive deficits associated with AD may be partly caused by dysfunction of cholinergic receptor subtype a7nAChR (a7nAChRs) in hippocampal neurons. a7nAChRs activation results in Ca<sup>2+</sup> influx and participate in the release of neurotransmitters;  $\alpha$ 7nAChRs also regulate neuronal excitability and LTP response, implicating a role for these receptors in neuronal function (108-110). Donepezil up-regulates nicotinic receptors in cortical neurons, this probably contributing to enhance neuroprotection(111). Furthermore, it has been suggested that AChEIs can activate reflex cholinergic anti-inflammatory pathway via  $\alpha$ 7nACh R one of the main mechanistic pathways by which this receptor exerts its anti-inflammatory efficacy isthrough the janus kinase2/ signal transducer and activator of transcription 3, as well as theirfeedback regulator suppressor of cytokine signaling 3 (JAK2/ STAT3/ SOCS3) signalingpathway; Such pathway plays a role in regulating IL-6 and the transcriptional factor nuclearfactor- kappa B (NF-κB)(112). One more study also explore that AChEIs also influence pro-inflammatory cytokines released from peripheral blood mononuclear cells, increasing oncostatin M, IL-1 $\beta$ , and IL-6 levels in AD patients after treatment(113). Other studies indicate that donepezil has a neuroprotective effect against oxygene-glucose deprivation injury and glutamate toxicity in cultured cortical neurons, and that this neuroprotection may be partially mediated by inhibition of the increase of intracellular calcium concentration(114). Previous study by singh et al.(95) demonstrating the relationship of donepezil and cholinergic system mediated increase calcium level reduce inflammatory mediators along with increase IGF and CREB involvement in reduced tau hyperphosphorylation leading neurogenesis in AD type dementia.

In summary, donepezil activates cholinergic subtype  $\alpha$ 7nAChR, further activate Gq type protein of Gprotein mediate enhance calcium level which could be at downstream activate PLC and CaMK mediated activation of SIK1/SIK3 responsible for suppression of AD. Additionally, PKA expression is also stimulated via CGRP that is directly associated with the administration of donepezil.

## **Conclusion:**

In conclusion, other factors such as aging, metal ion, virus, and microbiota may also contribute to AD pathogenesis via various mechanisms. Despite much knowledge that we have gained, no effective treatment strategies for AD have been successfully developed. Intervention for early-onset AD may require treatment at a young age, as calcium dependent Aβ aggregation and accumulation manifests early onset forms of the disease which further contribute in neuroinflammation and neurodegeneration in AD. Importantly, there are no drugs targeting SIKs that have been proven safe for clinical treatment for youths.Furthermore, future study warranted to provide new insights to therapeutic targets for treatment.

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## **References:**

1. Ferri CP, Prince M, Brayne C, Brodaty H, Fratiglioni L, Ganguli M, et al. Global prevalence of dementia: a Delphi consensus study. The Lancet. 2005;366(9503):2112-7.

2. Brookmeyer R, Johnson E, Ziegler-Graham K, Arrighi HM. Forecasting the global burden of Alzheimer's disease. Alzheimer's & dementia : the journal of the Alzheimer's Association. 2007;3(3):186-91.

3. Rizzi L, Rosset I, Roriz-Cruz M. Global Epidemiology of Dementia: Alzheimer's and Vascular Types. BioMed Research International. 2014;2014:908915.

4. 2012 Alzheimer's disease facts and figures. Alzheimer's & Dementia. 2012;8(2):131-68.

5. Winblad B, Amouyel P, Andrieu S, Ballard C, Brayne C, Brodaty H, et al. Defeating Alzheimer's disease and other dementias: a priority for European science and society. The Lancet Neurology. 2016;15(5):455-532.

6. Bellou V, Belbasis L, Tzoulaki I, Middleton LT, Ioannidis JPA, Evangelou E. Systematic evaluation of the associations between environmental risk factors and dementia: An umbrella review of systematic reviews and meta-analyses. Alzheimer's & dementia : the journal of the Alzheimer's Association. 2017;13(4):406-18.

7. Tarawneh R, Holtzman DM. The clinical problem of symptomatic Alzheimer disease and mild cognitive impairment. Cold Spring Harbor perspectives in medicine. 2012;2(5):a006148.

8. Zubenko GS, Zubenko WN, McPherson S, Spoor E, Marin DB, Farlow MR, et al. A collaborative study of the emergence and clinical features of the major depressive syndrome of Alzheimer's disease. The American journal of psychiatry. 2003;160(5):857-66.

9. Kalia M. Dysphagia and aspiration pneumonia in patients with Alzheimer's disease. Metabolism: clinical and experimental. 2003;52(10 Suppl 2):36-8.

10. Yiannopoulou KG, Papageorgiou SG. Current and future treatments for Alzheimer's disease. Therapeutic advances in neurological disorders. 2013;6(1):19-33.

11. Singh Y, Gupta G, Shrivastava B, Dahiya R, Tiwari J, Ashwathanarayana M, et al. Calcitonin generelated peptide (CGRP): A novel target for Alzheimer's disease. CNS neuroscience & therapeutics. 2017;23(6):457-61.

12. Weigand AJ, Bondi MW, Thomas KR, Campbell NL, Galasko DR, Salmon DP, et al. Association of anticholinergic medication and AD biomarkers with incidence of MCI among cognitively normal older adults. Neurology. 2020:10.1212/WNL.00000000010643.

13. Cummings JL, Morstorf T, Zhong K. Alzheimer's disease drug-development pipeline: few candidates, frequent failures. Alzheimer's research & therapy. 2014;6(4):37.

14. Godyń J, Jończyk J, Panek D, Malawska B. Therapeutic strategies for Alzheimer's disease in clinical trials. Pharmacological reports : PR. 2016;68(1):127-38.

15. Dansokho C, Heneka MT. Neuroinflammatory responses in Alzheimer's disease. Journal of Neural Transmission. 2018;125(5):771-9.

16. Traynelis SF, Wollmuth LP, McBain CJ, Menniti FS, Vance KM, Ogden KK, et al. Glutamate Receptor Ion Channels: Structure, Regulation, and Function. Pharmacological Reviews. 2010;62(3):405-

96.

17. Cull-Candy S, Brickley S, Farrant M. NMDA receptor subunits: diversity, development and disease. Current opinion in neurobiology. 2001;11(3):327-35.

18. Köhr G. NMDA receptor function: subunit composition versus spatial distribution. Cell and tissue research. 2006;326(2):439-46.

19. Parameshwaran K, Dhanasekaran M, Suppiramaniam V. Amyloid beta peptides and glutamatergic synaptic dysregulation. Experimental neurology. 2008;210(1):7-13.

20. Zhou Y, Danbolt N. GABA and Glutamate Transporters in Brain. Frontiers in Endocrinology. 2013;4(165).

21. Bell KF, Bennett DA, Cuello AC. Paradoxical upregulation of glutamatergic presynaptic boutons during mild cognitive impairment. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2007;27(40):10810-7.

22. Masliah E, Alford M, DeTeresa R, Mallory M, Hansen L. Deficient glutamate transport is associated with neurodegeneration in Alzheimer's disease. Annals of neurology. 1996;40(5):759-66.

23. Yang T, Li S, Xu H, Walsh DM, Selkoe DJ. Large Soluble Oligomers of Amyloid  $\beta$ -Protein from Alzheimer Brain Are Far Less Neuroactive Than the Smaller Oligomers to Which They Dissociate. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2017;37(1):152-63.

24. Jin M, Selkoe DJ. Systematic analysis of time-dependent neural effects of soluble amyloid  $\beta$  oligomers in culture and in vivo: Prevention by scyllo-inositol. Neurobiology of disease. 2015;82:152-63.

25. Hascup KN, Findley CA, Sime LN, Hascup ER. Hippocampal alterations in glutamatergic signaling during amyloid progression in AβPP/PS1 mice. Scientific Reports. 2020;10(1):14503.

26. Tapia-Rojas C, Cabezas-Opazo F, Deaton CA, Vergara EH, Johnson GVW, Quintanilla RA. It's all about tau. Progress in neurobiology. 2019;175:54-76.

27. Liu F, Grundke-Iqbal I, Iqbal K, Gong CX. Contributions of protein phosphatases PP1, PP2A, PP2B and PP5 to the regulation of tau phosphorylation. The European journal of neuroscience. 2005;22(8):1942-50.

28. Sontag E, Nunbhakdi-Craig V, Lee G, Bloom GS, Mumby MC. Regulation of the phosphorylation state and microtubule-binding activity of Tau by protein phosphatase 2A. Neuron. 1996;17(6):1201-7.

29. Drewes G, Mandelkow EM, Baumann K, Goris J, Merlevede W, Mandelkow E. Dephosphorylation of tau protein and Alzheimer paired helical filaments by calcineurin and phosphatase-2A. FEBS letters. 1993;336(3):425-32.

30. Vogelsberg-Ragaglia V, Schuck T, Trojanowski JQ, Lee VM. PP2A mRNA expression is quantitatively decreased in Alzheimer's disease hippocampus. Experimental neurology. 2001;168(2):402-12.

31. Gong CX, Singh TJ, Grundke-Iqbal I, Iqbal K. Phosphoprotein phosphatase activities in Alzheimer disease brain. Journal of neurochemistry. 1993;61(3):921-7.

32. Pei JJ, Tanaka T, Tung YC, Braak E, Iqbal K, Grundke-Iqbal I. Distribution, levels, and activity of glycogen synthase kinase-3 in the Alzheimer disease brain. Journal of neuropathology and experimental neurology. 1997;56(1):70-8.

33. Sontag E, Luangpirom A, Hladik C, Mudrak I, Ogris E, Speciale S, et al. Altered expression levels of the protein phosphatase 2A ABalphaC enzyme are associated with Alzheimer disease pathology. Journal of neuropathology and experimental neurology. 2004;63(4):287-301.

34. Tseng HC, Zhou Y, Shen Y, Tsai LH. A survey of Cdk5 activator p35 and p25 levels in Alzheimer's disease brains. FEBS letters. 2002;523(1-3):58-62.

35. Cacabelos R. Donepezil in Alzheimer's disease: From conventional trials to pharmacogenetics. Neuropsychiatr Dis Treat. 2007;3(3):303-33.

36. Tymianski M, Charlton MP, Carlen PL, Tator CH. Source specificity of early calcium neurotoxicity in cultured embryonic spinal neurons. The Journal of neuroscience : the official journal of the Society for Neuroscience. 1993;13(5):2085-104.

37. Choi DW, Koh JY, Peters S. Pharmacology of glutamate neurotoxicity in cortical cell culture: attenuation by NMDA antagonists. The Journal of neuroscience : the official journal of the Society for Neuroscience. 1988;8(1):185-96.

38. Koh JY, Choi DW. Selective blockade of non-NMDA receptors does not block rapidly triggered glutamate-induced neuronal death. Brain research. 1991;548(1-2):318-21.

39. Choi DW. Ionic dependence of glutamate neurotoxicity. The Journal of neuroscience : the official journal of the Society for Neuroscience. 1987;7(2):369-79.

40. Choi DW. Excitotoxic cell death. Journal of neurobiology. 1992;23(9):1261-76.

41. Danysz W, Parsons CG. The NMDA receptor antagonist memantine as a symptomatological and neuroprotective treatment for Alzheimer's disease: preclinical evidence. International journal of geriatric psychiatry. 2003;18(Suppl 1):S23-32.

42. Danysz W, Parsons CG, Mobius HJ, Stoffler A, Quack G. Neuroprotective and symptomatological action of memantine relevant for Alzheimer's disease--a unified glutamatergic hypothesis on the mechanism of action. Neurotoxicity research. 2000;2(2-3):85-97.

43. Wenk GL. Neuropathologic changes in Alzheimer's disease: potential targets for treatment. The Journal of clinical psychiatry. 2006;67 Suppl 3:3-7; quiz 23.

44. Mohamed A, Posse de Chaves E. A<i> $\beta$ </i> Internalization by Neurons and Glia. International Journal of Alzheimer&#x2019;s Disease. 2011;2011:127984.

45. Makitani K, Nakagawa S, Izumi Y, Akaike A, Kume T. Inhibitory effect of donepezil on bradykinininduced increase in the intracellular calcium concentration in cultured cortical astrocytes. Journal of Pharmacological Sciences. 2017;134(1):37-44.

46. Vélez-Fort M, Audinat E, Angulo MC. Functional alpha 7-containing nicotinic receptors of NG2-expressing cells in the hippocampus. Glia. 2009;57(10):1104-14.

47. Takarada T, Nakamichi N, Kawagoe H, Ogura M, Fukumori R, Nakazato R, et al. Possible neuroprotective property of nicotinic acetylcholine receptors in association with predominant upregulation of glial cell line-derived neurotrophic factor in astrocytes. Journal of neuroscience research. 2012;90(11):2074-85.

48. Wang X, Wang W, Li L, Perry G, Lee HG, Zhu X. Oxidative stress and mitochondrial dysfunction in Alzheimer's disease. Biochimica et biophysica acta. 2014;1842(8):1240-7.

49. Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G. Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. Biochimica et biophysica acta. 2010;1802(1):2-10.

50. Delekate A, Füchtemeier M, Schumacher T, Ulbrich C, Foddis M, Petzold GC. Metabotropic P2Y1 receptor signalling mediates astrocytic hyperactivity in vivo in an Alzheimer's disease mouse model. Nature Communications. 2014;5(1):5422.

51. Ronco V, Grolla AA, Glasnov TN, Canonico PL, Verkhratsky A, Genazzani AA, et al. Differential

deregulation of astrocytic calcium signalling by amyloid- $\beta$ , TNF $\alpha$ , IL-1 $\beta$  and LPS. Cell Calcium. 2014;55(4):219-29.

52. Grolla AA, Fakhfouri G, Balzaretti G, Marcello E, Gardoni F, Canonico PL, et al. Aβ leads to Ca2+ signaling alterations and transcriptional changes in glial cells. Neurobiology of Aging. 2013;34(2):511-22.

53. Pirttimaki TM, Codadu NK, Awni A, Pratik P, Nagel DA, Hill EJ, et al.  $\alpha$ 7 Nicotinic receptormediated astrocytic gliotransmitter release: A $\beta$  effects in a preclinical Alzheimer's mouse model. PloS one. 2013;8(11):e81828.

54. Talantova M, Sanz-Blasco S, Zhang X, Xia P, Akhtar MW, Okamoto S, et al. Aβ induces astrocytic glutamate release, extrasynaptic NMDA receptor activation, and synaptic loss. Proceedings of the National Academy of Sciences of the United States of America. 2013;110(27):E2518-27.

55. Benilova I, De Strooper B. Neuroscience. Promiscuous Alzheimer's amyloid: yet another partner. Science (New York, NY). 2013;341(6152):1354-5.

56. Nicolaides NC, Kyratzi E, Lamprokostopoulou A, Chrousos GP, Charmandari E. Stress, the stress system and the role of glucocorticoids. Neuroimmunomodulation. 2015;22(1-2):6-19.

57. Smith SM, Vale WW. The role of the hypothalamic-pituitary-adrenal axis in neuroendocrine responses to stress. Dialogues in clinical neuroscience. 2006;8(4):383-95.

58. Ellis BJ, Del Giudice M. Beyond allostatic load: rethinking the role of stress in regulating human development. Development and psychopathology. 2014;26(1):1-20.

59. Sapolsky RM, Krey LC, McEwen BS. Prolonged glucocorticoid exposure reduces hippocampal neuron number: implications for aging. The Journal of neuroscience : the official journal of the Society for Neuroscience. 1985;5(5):1222-7.

60. Terrill-Usery SE, Mohan MJ, Nichols MR. Amyloid- $\beta$ (1-42) protofibrils stimulate a quantum of secreted IL-1 $\beta$  despite significant intracellular IL-1 $\beta$  accumulation in microglia. Biochimica et biophysica acta. 2014;1842(11):2276-85.

61. Gratuze M, Leyns CEG, Holtzman DM. New insights into the role of TREM2 in Alzheimer's disease. Molecular neurodegeneration. 2018;13(1):66.

62. Kielian T. Toll-like receptors in central nervous system glial inflammation and homeostasis. Journal of neuroscience research. 2006;83(5):711-30.

63. Olson JK, Miller SD. Microglia initiate central nervous system innate and adaptive immune responses through multiple TLRs. Journal of immunology (Baltimore, Md : 1950). 2004;173(6):3916-24.

64. Fuller JP, Stavenhagen JB, Teeling JL. New roles for Fc receptors in neurodegeneration-the impact on Immunotherapy for Alzheimer's Disease. Front Neurosci. 2014;8:235-.

65. Husemann J, Loike JD, Kodama T, Silverstein SC. Scavenger receptor class B type I (SR-BI) mediates adhesion of neonatal murine microglia to fibrillar beta-amyloid. Journal of neuroimmunology. 2001;114(1-2):142-50.

66. Moore KJ, El Khoury J, Medeiros LA, Terada K, Geula C, Luster AD, et al. A CD36-initiated signaling cascade mediates inflammatory effects of beta-amyloid. The Journal of biological chemistry. 2002;277(49):47373-9.

67. Lue LF, Walker DG, Brachova L, Beach TG, Rogers J, Schmidt AM, et al. Involvement of microglial receptor for advanced glycation endproducts (RAGE) in Alzheimer's disease: identification of a cellular activation mechanism. Experimental neurology. 2001;171(1):29-45.

68. Landreth GE, Reed-Geaghan EG. Toll-like receptors in Alzheimer's disease. Current topics in

microbiology and immunology. 2009;336:137-53.

69. Fassbender K, Walter S, Kühl S, Landmann R, Ishii K, Bertsch T, et al. The LPS receptor (CD14) links innate immunity with Alzheimer's disease. FASEB journal : official publication of the Federation of American Societies for Experimental Biology. 2004;18(1):203-5.

70. Li R, Yang L, Lindholm K, Konishi Y, Yue X, Hampel H, et al. Tumor necrosis factor death receptor signaling cascade is required for amyloid-beta protein-induced neuron death. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2004;24(7):1760-71.

71. Molina-Holgado E, Ortiz S, Molina-Holgado F, Guaza C. Induction of COX-2 and PGE(2) biosynthesis by IL-1beta is mediated by PKC and mitogen-activated protein kinases in murine astrocytes. British journal of pharmacology. 2000;131(1):152-9.

72. Pan XD, Zhu YG, Lin N, Zhang J, Ye QY, Huang HP, et al. Microglial phagocytosis induced by fibrillar  $\beta$ -amyloid is attenuated by oligomeric  $\beta$ -amyloid: implications for Alzheimer's disease. Molecular neurodegeneration. 2011;6:45.

73. Cregg JM, DePaul MA, Filous AR, Lang BT, Tran A, Silver J. Functional regeneration beyond the glial scar. Experimental neurology. 2014;253:197-207.

74. Parpura V, Heneka MT, Montana V, Oliet SH, Schousboe A, Haydon PG, et al. Glial cells in (patho)physiology. Journal of neurochemistry. 2012;121(1):4-27.

75. Scimemi A, Meabon JS, Woltjer RL, Sullivan JM, Diamond JS, Cook DG. Amyloid- $\beta$ 1-42 slows clearance of synaptically released glutamate by mislocalizing astrocytic GLT-1. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2013;33(12):5312-8.

76. Wang WY, Tan MS, Yu JT, Tan L. Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease. Annals of translational medicine. 2015;3(10):136.

77. Wang L, Jiang Q, Chu J, Lin L, Li XG, Chai GS, et al. Expression of Tau40 induces activation of cultured rat microglial cells. PloS one. 2013;8(10):e76057.

78. Ma L, Sun P, Zhang JC, Zhang Q, Yao SL. Proinflammatory effects of S100A8/A9 via TLR4 and RAGE signaling pathways in BV-2 microglial cells. International journal of molecular medicine. 2017;40(1):31-8.

79. Heneka MT, Kummer MP, Stutz A, Delekate A, Schwartz S, Vieira-Saecker A, et al. NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice. Nature. 2013;493(7434):674-8.

80. Halle A, Hornung V, Petzold GC, Stewart CR, Monks BG, Reinheckel T, et al. The NALP3 inflammasome is involved in the innate immune response to amyloid-beta. Nature immunology. 2008;9(8):857-65.

81. Wang Z, Takemori H, Halder SK, Nonaka Y, Okamoto M. Cloning of a novel kinase (SIK) of the SNF1/AMPK family from high salt diet-treated rat adrenal. FEBS letters. 1999;453(1-2):135-9.

82. Taub M, Springate JE, Cutuli F. Targeting of renal proximal tubule Na,K-ATPase by salt-inducible kinase. Biochemical and biophysical research communications. 2010;393(3):339-44.

83. Chen F, Chen L, Qin Q, Sun X. Salt-Inducible Kinase 2: An Oncogenic Signal Transmitter and Potential Target for Cancer Therapy. Frontiers in oncology. 2019;9:18.

84. Hashimoto YK, Satoh T, Okamoto M, Takemori H. Importance of autophosphorylation at Ser186 in the A-loop of salt inducible kinase 1 for its sustained kinase activity. Journal of cellular biochemistry. 2008;104(5):1724-39.

85. Katoh Y, Takemori H, Horike N, Doi J, Muraoka M, Min L, et al. Salt-inducible kinase (SIK) isoforms: their involvement in steroidogenesis and adipogenesis. Molecular and cellular endocrinology. 2004;217(1-2):109-12.

86. Feldman JD, Vician L, Crispino M, Hoe W, Baudry M, Herschman HR. The salt-inducible kinase, SIK, is induced by depolarization in brain. Journal of neurochemistry. 2000;74(6):2227-38.

87. Horike N, Takemori H, Katoh Y, Doi J, Min L, Asano T, et al. Adipose-specific expression, phosphorylation of Ser794 in insulin receptor substrate-1, and activation in diabetic animals of salt-inducible kinase-2. The Journal of biological chemistry. 2003;278(20):18440-7.

88. Zhang Y, Gao W, Yang K, Tao H, Yang H. Salt-Inducible Kinase 1 (SIK1) is Induced by Alcohol and Suppresses Microglia Inflammation via NF-κB Signaling. Cellular Physiology and Biochemistry. 2018;47(4):1411-21.

89. Yong Kim S, Jeong S, Chah K-H, Jung E, Baek K-H, Kim S-T, et al. Salt-Inducible Kinases 1 and 3 Negatively Regulate Toll-Like Receptor 4-Mediated Signal. Molecular Endocrinology. 2013;27(11):1958-68.

90. Sanosaka M, Fujimoto M, Ohkawara T, Nagatake T, Itoh Y, Kagawa M, et al. Salt-inducible kinase 3 deficiency exacerbates lipopolysaccharide-induced endotoxin shock accompanied by increased levels of pro-inflammatory molecules in mice. Immunology. 2015;145(2):268-78.

91. Yong Kim S, Jeong S, Chah KH, Jung E, Baek KH, Kim ST, et al. Salt-inducible kinases 1 and 3 negatively regulate Toll-like receptor 4-mediated signal. Molecular endocrinology (Baltimore, Md). 2013;27(11):1958-68.

92. Sasaki T, Takemori H, Yagita Y, Terasaki Y, Uebi T, Horike N, et al. SIK2 Is a Key Regulator for Neuronal Survival after Ischemia via TORC1-CREB. Neuron. 2011;69(1):106-19.

93. Ma L, Manaenko A, Ou Y-B, Shao A-W, Yang S-X, Zhang JH. Bosutinib Attenuates Inflammation via Inhibiting Salt-Inducible Kinases in Experimental Model of Intracerebral Hemorrhage on Mice. Stroke. 2017;48(11):3108-16.

94. Xu BE, Stippec S, Lazrak A, Huang CL, Cobb MH. WNK1 activates SGK1 by a phosphatidylinositol 3-kinase-dependent and non-catalytic mechanism. The Journal of biological chemistry. 2005;280(40):34218-23.

95. Singh Y, Gupta G, Shrivastava B, Dahiya R, Tiwari J, Ashwathanarayana M, et al. Calcitonin generelated peptide (CGRP): A novel target for Alzheimer's disease. CNS neuroscience & therapeutics. 2017;23(6):457-61.

96. Bertorello AM, Zhu J-K. SIK1/SOS2 networks: decoding sodium signals via calcium-responsive protein kinase pathways. Pflügers Archiv - European Journal of Physiology. 2009;458(3):613.

97. Sakamoto K, Bultot L, Göransson O. The Salt-Inducible Kinases: Emerging Metabolic Regulators. Trends in endocrinology and metabolism: TEM. 2018;29(12):827-40.

98. Patel K, Foretz M, Marion A, Campbell DG, Gourlay R, Boudaba N, et al. The LKB1-salt-inducible kinase pathway functions as a key gluconeogenic suppressor in the liver. Nat Commun. 2014;5:4535.

99. Sjöström M, Stenström K, Eneling K, Zwiller J, Katz AI, Takemori H, et al. SIK1 is part of a cell sodium-sensing network that regulates active sodium transport through a calcium-dependent process. Proceedings of the National Academy of Sciences of the United States of America. 2007;104(43):16922-7.

100. Suzuki T, Imai J, Yamada T, Ishigaki Y, Kaneko K, Uno K, et al. Interleukin-6 enhances glucose-

stimulated insulin secretion from pancreatic beta-cells: potential involvement of the PLC-IP3-dependent pathway. Diabetes. 2011;60(2):537-47.

101. Sasaki T, Takemori H, Yagita Y, Terasaki Y, Uebi T, Horike N, et al. SIK2 is a key regulator for neuronal survival after ischemia via TORC1-CREB. Neuron. 2011;69(1):106-19.

102. Koo SH, Flechner L, Qi L, Zhang X, Screaton RA, Jeffries S, et al. The CREB coactivator TORC2 is a key regulator of fasting glucose metabolism. Nature. 2005;437(7062):1109-11.

103. Nixon M, Stewart-Fitzgibbon R, Fu J, Akhmedov D, Rajendran K, Mendoza-Rodriguez MG, et al. Skeletal muscle salt inducible kinase 1 promotes insulin resistance in obesity. Molecular metabolism. 2016;5(1):34-46.

104. Winkler EA, Nishida Y, Sagare AP, Rege SV, Bell RD, Perlmutter D, et al. GLUT1 reductions exacerbate Alzheimer's disease vasculo-neuronal dysfunction and degeneration. Nat Neurosci. 2015;18(4):521-30.

105. Choi S, Kim W, Chung J. Drosophila salt-inducible kinase (SIK) regulates starvation resistance through cAMP-response element-binding protein (CREB)-regulated transcription coactivator (CRTC). The Journal of biological chemistry. 2011;286(4):2658-64.

106. Uebi T, Itoh Y, Hatano O, Kumagai A, Sanosaka M, Sasaki T, et al. Involvement of SIK3 in glucose and lipid homeostasis in mice. PloS one. 2012;7(5):e37803.

107. Zhao J, Zhang X, Gao T, Wang S, Hou Y, Yuan P, et al. SIK2 enhances synthesis of fatty acid and cholesterol in ovarian cancer cells and tumor growth through PI3K/Akt signaling pathway. Cell death & disease. 2020;11(1):25.

108. Dani JA. Nicotinic receptor activity alters synaptic plasticity. TheScientificWorldJournal. 2001;1:393-5.

109. Gray R, Rajan AS, Radcliffe KA, Yakehiro M, Dani JA. Hippocampal synaptic transmission enhanced by low concentrations of nicotine. Nature. 1996;383(6602):713-6.

110. Ji D, Lape R, Dani JA. Timing and location of nicotinic activity enhances or depresses hippocampal synaptic plasticity. Neuron. 2001;31(1):131-41.

111. Kume T, Sugimoto M, Takada Y, Yamaguchi T, Yonezawa A, Katsuki H, et al. Up-regulation of nicotinic acetylcholine receptors by central-type acetylcholinesterase inhibitors in rat cortical neurons. European journal of pharmacology. 2005;527(1-3):77-85.

112. Wazea SA, Wadie W, Bahgat AK, El-Abhar HS. Galantamine anti-colitic effect: Role of alpha-7 nicotinic acetylcholine receptor in modulating Jak/STAT3, NF-κB/HMGB1/RAGE and p-AKT/Bcl-2 pathways. Scientific Reports. 2018;8(1):5110.

113. Reale M, Iarlori C, Gambi F, Lucci I, Salvatore M, Gambi D. Acetylcholinesterase inhibitors effects on oncostatin-M, interleukin-1 beta and interleukin-6 release from lymphocytes of Alzheimer's disease patients. Experimental gerontology. 2005;40(3):165-71.

114. Akasofu S, Kimura M, Kosasa T, Ogura H, Sawada K. Protective effect of donepezil in primarycultured rat cortical neurons exposed to N-methyl-d-aspartate (NMDA) toxicity. European journal of pharmacology. 2006;530(3):215-22.