

Chemical Composition Of Essential Oils From *Rhzyastricta* And Its Antioxidant Activity

Awais Madni¹, Muhammad Adnan Iqbal^{1*}, Rushina Batool¹, Athar Mahmood^{2*}, Maria Naqve³, Muhammad Mansoor Javaid³, Muhammad Hasnain¹, Mehwish Naseer⁵, Muhammad Awais Akram¹

¹Department of Chemistry, University of Agriculture, Faisalabad 38040, Pakistan

²Department of Agronomy, University of Agriculture, Faisalabad 38040, Pakistan

³Department of Boatany, University of Agriculture, Faisalabad 38040, Pakistan

⁴Department of Agronomy, College of Agriculture, University of Sargodha, Sargodha, 40100, Pakistan

⁵Department of Boatany, Government College Women University Faisalabad, Faisalabad, 38040, Pakistan

Abstract

Rhzyastricta is a medicinal plant. It is used for the treatment of many diseases such as Diabetes, Coating infections, cancer and inflammations. The chemical constituents of *R. stricta* essential oils were analyzed by using GC-FID. Seven components were analyzed from essential oils of *R. stricta*. These chemical components were Acetyl Pyridine (0.04%), Pyrazine (0.10%), Lavandulol (0.03%), Pinocarveol (0.02%), Hexadienolisobutanoate (0.14%), Silphiperfol-4,7 (14)-diene (0.04%) and Cubenol (1.22%). Total phenolic contents and total flavonoid contents of essential oils of *R. stricta* were also assessed and were found in the range of 13.5 ± 0.3 (mg GAE/g) and 3.53 ± 0.40 (mg QE/g). The antioxidant activity of essential oils of *R. stricta* was analyzed by using DPPH free radical scavenging assay. The inhibition % for DPPH was 23.33 ± 0.15 $\mu\text{g/mL}$ which was done by essential oils of *R. stricta*. Chemical composition of essential oils of *R. stricta* was determined for the first time.

Keywords: *Rhzyastricta*, essential oils, GC-FID, total phenolics, total flavonoids

Introduction

Medicinal plants are used in herbal medicines and also have greater medicinal efficacy. These medicinal vegetations have constituents which are used for progress and amalgamation of drugs. These medicinal plants show key part in the expansion of human belief all over the world. Some vegetations deliberate as significance base of diet and due to this reason these vegetations suggested for their beneficial standards (Rasool Hassan, 2012). Plant kingdom has one of the greatest family which is known as Apocynaceae. Apocynaceae family comprise of genera which are 424 and species which are 4600. Plants which belongs to Apocynaceae family are inherent in Pakistan, Sri Lanka, India, China and Bangladesh (Bhadane, Patil, Maheshwari, & Patil, 2018). *Rhzyastricta* common name in Pakistan is "Sahwar" and it

is called “Izrushk” in balochi language in Pakistan. Its Arabic name is “Harmal” and its Persian name is “Eshvarak” (Marwat, Usman, Shah, Anwar, & Ullah, 2012). Rhazya is inherent to Asia to Southwest and it was designated in the year of 1835 as genus. Two types of persistent basils and bushes present in it. It is dispersed all over the zones which are subtropics specifically in Pakistan and India which belongs to South Asia and Arabian Peninsula which belongs to Central East (Abdul-Hameed et al., 2021). Two types of species which are known as *R. orientalis* and *R. stricta* included in this genus and this genus belongs to family Apocynaceae. *R. stricta* is small perennial woody shrubby plant which is also a poisonous plant and it is also vertical and exposed (Akhgari et al., 2015). The tallness of *R. stricta* is 90 cm. Its branches densified, leaves are alternative and stem is smoothy. With passage of time the color of leaves changed to yellow. Its seeds are small aerial. Its fruit are light yellowish (Marwat et al., 2012). *R. stricta* a small goblin bush which is generally dispersed in different areas of Pakistan such as Khyber, Balochistan, NWFP, Karachi, Sindh, Dargai, grasslands which are present among Jhelum and Indus and salty series. The leaves have vertical shells (Sultana & Khalid, 2010). *R. stricta* has many branches which are compact and these branches climbing from foundation and also evergreen. *R. stricta* is basically firm, vertical climber. Its branches majorly erected from the starting point and its stem is horizontal and central (Akyalcin, Ozen, & Dulger, 2006).

Its leaves have smoothy exterior. They are 12 cm above the dwarf trunk, 10 cm extensive and 1.5 cm exteriorly. The nature of leaves is fibrous and they have alternative edge which elongated near the starting point. They also have a midrib which is protruding. They have severe top. Leaves are also oval and rigid. They are sedentary and vertical from whole edges. Its flowers length is 2.5 cm and their color are white and they are present close to spike of twigs. Flowers are wing shaped and ambisexual. They have small trunk in which stamens are injected. The color of petal is white. The length of corolla is for about 1.4 cm and 4mm for calyx. The color of corolla is also white. The lobes are intensively critical from three sided. The lobes are 15 mm in length and they have a tube which is greenish brown and this tube extended overhead the center. The lobes are basically oval and their top part is curved. Their length is greater than tray like branch. The lobes which are present in appendages are bluish from backside and they are white from inside. They are also thick headed. The stigma is annular. The filaments are small and thread like. Three types of stomata are present in *R. stricta* which are irregular, unequal and parallel (Bukhari, Al-Otaibi, & Ibbrahim, 2017).

Table 1. Fragments of *Rhazyastricta* which are used for different disorders

Disorders	Fragment Used
Diabetes	Entire plant
Coating infections	Ovary, Greeneries
Cancer	Greeneries
Injuries	Entire plant
Inflammations and spots problem of face	Greeneries
Helminthiasis	Entire plant

Plant *R. stricta* is mainly practiced for curing different kinds of illnesses in Iran, Qatar, Iraq, Afghanistan, Pakistan and India. It is practiced for medication of cancer, tumor and diabetes. Different fragments obtained from it used as antioxidant, antiviral, antifungal, antibacterial, antimutagenic, anti-inflammatory. Folk medication also practices it for treatment of pain occurring in stomach, rashes occurring on skin, sore occurring in throat and for eyes swelling (Lanjwani, Ganghro, & Khuhawar, 2018). *R. stricta* examined as chief medicinal plant practice as medication in conventional drugs. And they contribute key part for curing animal and humanoid illnesses (Al-Hasawi & Al-Harbi, 2014). *R. stricta* actual practice is defined in outmoded medication had ascribed due to existence of alkaloids which are called as indole. When *R. stricta* was examined than it was identified that constituents which are alkaloid have greater biologic actions (A. I. Elkady, 2013). *R. stricta* which is chief medication plant show anti-cancerous and anti-oxidant qualities and also have free radical foraging belongings and used in old style medication. The research was concluded to discover the anti-cancerous efficacy of alkaloids which are obtained from *R. stricta* for cancer cell line which is A549. Crude alkaloid extract of *Rhazya stricta* remarkably improve ability of cisplatin as anti-proliferative and repress the development of cancer cell line A549 (A. I. Elkady, Hussein, & Abu-Zinadah, 2014). Distillate which obtained from *R. stricta* used for curing different disorders such as rheumatism, helminthiasis and diabetes. Further, it was described that crude distillate gained from *R. stricta* cause inhibition of propagation of cells and persuade the death of cells of apoptosis in cancerous lines such as MB-231 and MCF-7. More than 100 different kinds of alkaloids are separated, categorized and recognized from the leaves, stems and roots of *R. stricta*. The reality is that alkaloids which isolated from *R. stricta* are chief significant phytochemicals which are recognized due to anti-metastatic and anti-proliferation potentials on several kinds of cancerous cells in-vivo and in-vitro respectively (Lu, Bao, Chen, Huang, & Wang, 2012). The previous work on *R. stricta* is available on plant extraction but the current research work is done for extraction of EOs from *R. stricta*. The main objective to conduct present research was to extract EOs from *R. stricta*, to characterize EOs by using GC-FID and to check antioxidant activity of EOs.

Materials and Methods

Collection and identification of plant material

The plant sample was collected from hilly areas of District D.G Khan, Punjab, Pakistan. The plant sample was collected from this area in the month of March. The plant sample was identified by botanist (Dr. Mansoor Hameed), University of Agriculture, Faisalabad. The research work was done at Organometallic and Coordination Chemistry Laboratory of Postgraduate Agriculture Research Station (PARS), University of Agriculture Faisalabad.

Chemical and reagents

n-hexane (DAEJUNG, Korea), Distilled water and Celite (DAEJUNG, Korea) were used for extraction of essential oils. And for biological activities Folin-Ciocalteu (FC) reagent, Sodium carbonate (SIGMA-ALDRICH, Germany), Sodium nitrate (SIGMA-ALDRICH, Germany), Aluminum chloride (SIGMA-ALDRICH, Germany) and DPPH solution were used.

Extraction of essential oils

Hydro-distillation method was basically used for extraction of essential oils from *R. stricta*. First of all, collection of plant sample was done. The plant sample was washed with the help of distilled water. After it plant sample was dried for some time to evaporate water. The plant sample was raptured slightly with help of pestle and mortar and was weighted with the help of analytical balance. Total plant sample was for about 8560g. Distillery was ON and water supply was also started in distillery and after sometime distillery was filled up with water. A flask of about 500 mL was linked with distillery for collection of water and EOs. First batch of for about 2140g was put in distillery containing water and its led was covered. After sometime distillery was boiled up and finally boiling of distillery water along with EOs passing through pipe which linked with distillery and one end of pipe is in the flask in which water and EOs start collected. Drop by drop water and EOs start collected. The upper layer was EOs layer and lower layer was water layer. When flask filled up with water and EOs lower water layer was removed from flask upper layer of EOs remains as it was earlier. This process continues for about 2 hours than after it first batch of plant sample was removed and second batch of plant sample was added in the same way. Same procedure was repeated for third and fourth batch of plant sample. This procedure was continued for about 8 to 9 hours. Lower water layer was removed and upper EOs layer remains as it was before and become thick was passage of time. When whole procedure was completed than water was completely removed from flask than some quantity of n-hexane was added in flask containing EOs. So EOs which present on walls of flask moved in n-hexane. Finally, n-hexane containing EOs was passed through five-layer filter paper which is present in funnel filled with celite for removal of water so that pure EOs obtained. After sometime whole sample passed through 5-layered filter paper and pure EOs was obtained. The pure EOs was put in 5mL glass vails(Elyemni et al., 2019).

Characterization of essential oils

GC majorly used for volatile constituent's analysis. And an FID is systematic apparatus that calculate analytical sample in gaseous streamlet. Detector FID commonly used in GC. The method through which concluding information exhibited and it mainly relays on software and computer. Mostly FID is linked to GC arrangement. The EOs GC-FID examination was achieved with GC (Perkin Elmer) with Clarus (480) which is fortified with detector such as FID and a tube column like Elite-5 (PerkinElmer; 35 m × 0.30 mm × 0.30 μm). The temperature of oven was automated. The temperature of injector and detector were 220°C and 280°C, correspondingly. The (99.99%) helium at 0.6 (mL min⁻¹) drift rate was used as carrier gas. The EOs sample of for about 2 μL were inoculated with help of split mode. The EOs % age composition was determined by means of retention time and peak area(Silva-Flores et al., 2019).

Antioxidant activity

Total phenolic contents (TPC)

2μLFolin-Ciocalteu (FC) reagent was taken in culture tube. Then 800μL of Na₂CO₃ was added in culture tube containing FC reagent. After it 1mL of essential oil sample was added in culture tube containing FC reagent and Na₂CO₃. This culture tube was left for two hours. After two hours small amount was taken from this culture tube and added on ELISA plate. This ELISA plate was placed in spectrophotometer and reading was taken on 765nm wavelength. The calculation of total phenolic contents was done with help

of gallic acid calibration curve. The outcomes were represented as gallic acid equivalence (GAE) per dry weight. The experiment was repeated in a triplicate way (Anwar, Ali, Hussain, & Shahid, 2009).

Total flavonoid contents (TFC)

1.25mL distilled water was taken in culture tube. Then 250 μ L essential oil sample was added culture tube containing distilled water. Then 75 μ L of NaNO₂ was added in culture tube which contain distilled water and essential oil sample. After it 150 μ L of AlCl₃ was added in culture tube which already contain distilled water, essential oil sample and NaNO₂. It was left for five minutes. After five minutes 250 μ L amount from culture tube was taken and added on ELISA plate. Then this ELISA plate was placed in spectrophotometer and reading was taken on 510nm wavelength. Three readings were taken at 510 nm wavelength. The total flavonoid contents were represented as quercetin equivalent (QE) per gram of dry weight. The experiment was repeated three times(Riaz et al., 2012).

DPPH radical scavenging assay

Firstly, 1mL of DPPH solution was taken in Eppendorf tube. Then 10 μ L of essential oil sample was added in Eppendorf tube containing DPPH solution. Eppendorf tube containing DPPH solution and sample was left for half hour in dark place because it was light sensitive. After half hour Eppendorf tube containing DPPH solution and sample was taken. 250 μ L amount from Eppendorf tube was taken and added on ELISA plate. Then ELISA plate was placed in spectrophotometer and reading was taken on 510nm wavelength. Three readings of absorbance of blank and sample solution were taken at 510nm wavelength. The % inhibition of DPPH was calculated by this formula.

$$\text{DPPH inhibition (A. I. J. G. Elkady \& biology)} = 100 \times (A_{\text{blank}} - A_{\text{sample}} / A_{\text{blank}})$$

A_{blank} is the absorbance of control reaction mixture without including the test of blank solution compounds and A_{sample} is the absorbance of sample solution (Abdullah I Hussain, Anwar, Shahid, Ashraf, & Przybylski, 2010).

Results and discussion

Chemical composition of EOs of *R. stricta*

The first peak at retention time 1.96 (minute) is the peak of solvent (n-Hexane, 98.41%) which is used for the extraction of EOs from *R. stricta*. The second peak with retention time 8.84 (minute) is the peak of Acetyl Pyridine. Third peak with retention time 11.23 (minute) is the peak of Pyrazine. Forth peak with retention time 13.95 (minute) is the peak of Lavandulol. Fifth peak with retention time 15.00 (minute) is the peak of Pinocarveol. Sixth peak with retention time 15.92 (minute) is the peak of Hexadienoliso but a noate. Seventh peak with retention time 22.77 (minute) is the peak of Silphiperfol-4,7 (14)-diene. Last eighth peak with retention time 34.65 (minute) is the peak of Cubenol. So identified compounds by GC-FID are Acetyl Pyridine (0.04%), Pyrazine (0.10%), Lavandulol (0.03%), Pinocarveol (0.02%), Hexadienoliso but a noate (0.14%), Silphiperfol-4,7 (14)-diene (0.04%) and Cubenol (1.22%) which are presented in (Table 2) and the structures of all these identified compounds are shown in (Figure 2)(Adams, 2007). The chromatogram of EOs of *R. stricta* is shown in (Figure 1).

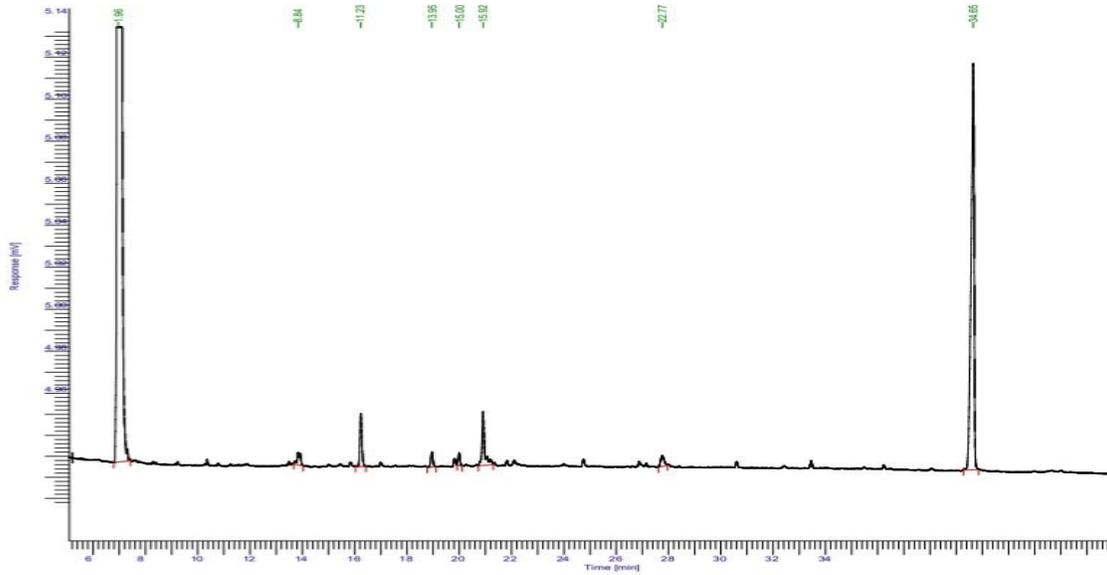
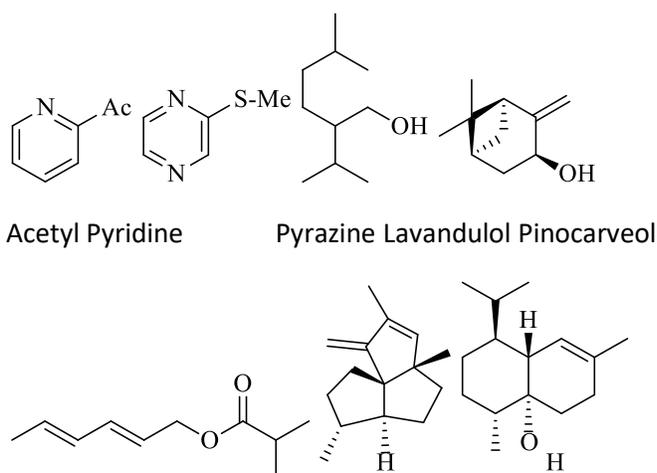


Figure 1. GC-FID chromatogram of *R. stricta* EOs

Table 2. *R. stricta* EOs identified compounds by GC-FID

Peak No	Retention Time	Area	Compound	Formula
1	1.96	98.41	n-Hexane (Solvent)	C ₆ H ₁₂
2	8.84	0.04	Acetyl Pyridine	C ₇ H ₇ NO
3	11.23	0.10	Pyrazine	C ₅ H ₆ N ₂ S
4	13.95	0.03	Lavandulol	C ₁₀ H ₂₂ O

5	15.00	0.02	Pinocarveol	C ₁₀ H ₁₆ O
6	15.92	0.14	Hexadienolisobutanoate	C ₁₀ H ₁₆ O ₂
7	22.77	0.04	Silphiperfol-4,7 (14)-diene	C ₁₅ H ₂₂
8	34.65	1.22	Cubanol	C ₁₅ H ₂₆ O



Hexadienolisobutanoate Silphiperfol-4,7 (14)-diene Cubanol

Figure 2. Structures of compounds identified by GC-FID

Total phenolic and total flavonoid contents

The total phenolic contents and total flavonoid contents were 13.5 ± 0.3 (mg GAE/g) of dry matter and 3.53 ± 0.40 (mg QE/g) of dry matter. Total phenolic contents were determined by Folin-Ciocalteu (FC) process. Gallic acid was taken as a standard for determination of total phenolic contents. Quercetin was taken as a standard for determination of total flavonoid contents. The total phenolic contents were greater as compared to total flavonoid contents in the EOs of *R. stricta*. The total phenolic contents have redox properties which are accountable for activity as antioxidant (Tohidi, Rahimmalek, & Arzani, 2017). The total phenolic and flavonoid contents of EOs of *R. stricta* are shown in (Table 3) and total phenolic and flavonoid contents are shown in (Figure 3).

Table 3. Total phenolic contents and Total flavonoid contents of *R. stricta* EOs

Rhazyastriacta EOs	TPC (mg GAE/g) of dry matter	TFC (mg QE/g) of dry matter
	13.5 ± 0.3	3.53 ± 0.40

The results are represented as mean \pm SD of triplicate experiments

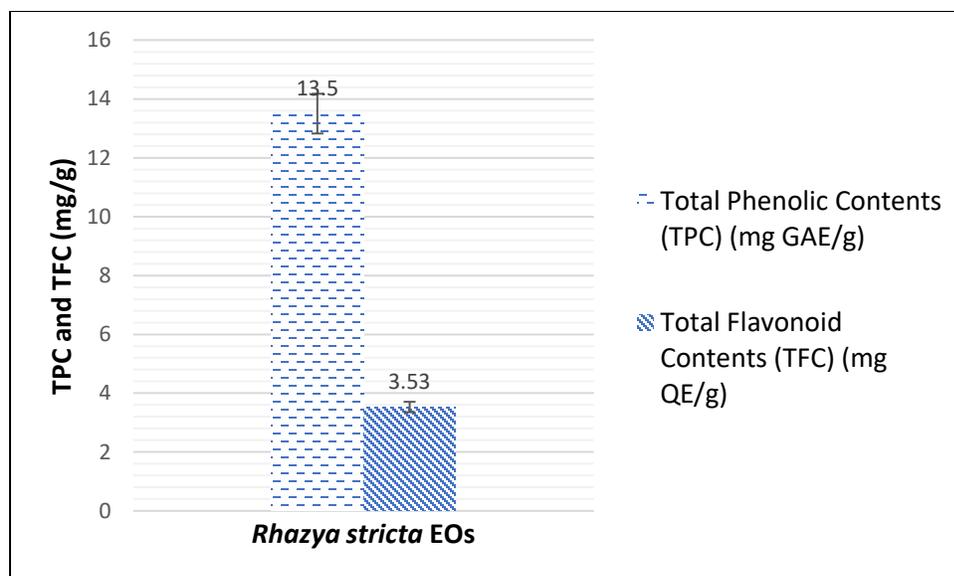


Figure 3. Total phenolic contents and total flavonoid contents of EOs

of *R. stricta*

DPPH radical scavenging assay

DPPH standard solution was used to determine antioxidant activity of EOs of *R. stricta*. DPPH solution has purple color. The EOs of *R. stricta* changed the purple color of DPPH into yellow color. The stable color of DPPH was purple. Due to receiving of proton from EOs the purple color of DPPH changed into yellow color. The inhibition % was 23.33 ± 0.15 $\mu\text{g/mL}$ which was done by essential oils of *Rhazya stricta* for DPPH free radical scavenging activity (Abdullah Ijaz Hussain, Anwar, Sherazi, & Przybylski, 2008). Antioxidant activity of EOs of *R. stricta* for DPPH assay is shown in (Table 4) and DPPH inhibition done by EOs of *R. stricta* is shown by bar graph in (Figure 4).

Table 4. Antioxidant activity of EOs of *R. stricta* for DPPH assay

Rhazya stricta EOs	DPPH inhibition $\mu\text{g/mL}$
	23.33 ± 0.15

The result is represented as mean \pm SD of triplicate experiments

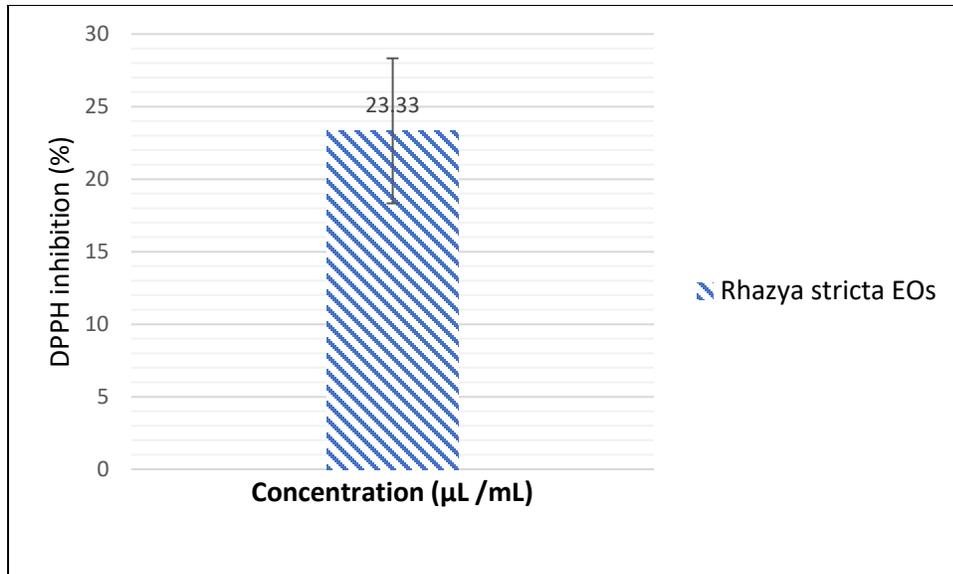


Figure 4. DPPH inhibition done by R. stricta EOs

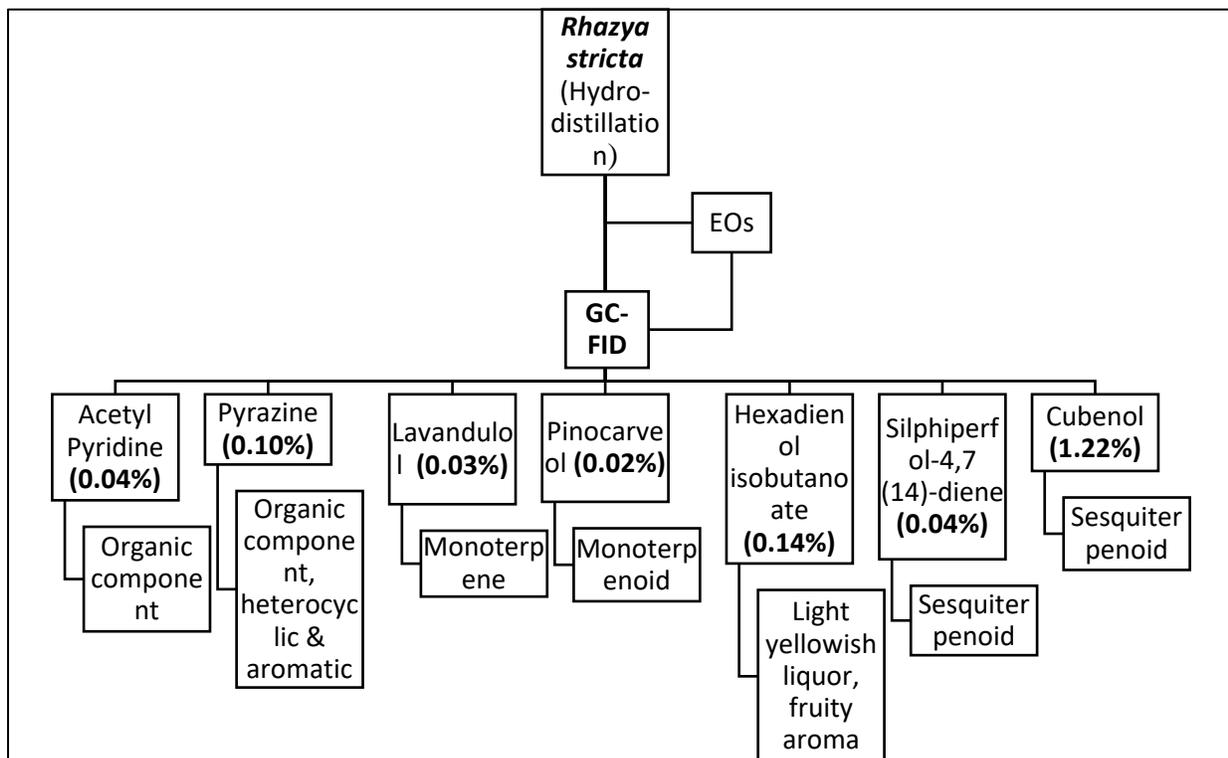


Figure 5. Schematic representation of characterization of R. stricta EOs by GC-FID

Schematic representation of characterization of EOs of R. stricta by GC-FID is shown in (Figure 5). The first compound which is identified by GC-FID is Acetyl Pyridine which is organic component. The second compound is Pyrazine which is also organic component. Third compound is Lavandulol which is monoterpene. Fourth compound is Pinocarveol which is monoterpenoid. Fifth compound is Hexadienoliso

but an oate which is light yellowish liquor with fruity aroma. Sixth compound is Silphiperfol-4,7 (14)-diene which is sesquiterpenoid. The last compound is Cubenol which is also sesquiterpenoid(Adams, 2007).

Conclusion

The chemical composition of essential oils of *R. stricta* is evaluated for first time. The chemical composition examination was done by using GC-FID technique. Seven components were identified by GC-FID. These components are Acetyl Pyridine (0.04%), Pyrazine (0.10%), Lavandulol (0.03%), Pinocarveol (0.02%), Hexadienoliso but a noate (0.14%), Silphiperfol-4,7 (14)-diene (0.04%) and Cubenol (1.22%). Total phenolic and flavonoid compounds which obtained from *R. stricta* were 13.5 ± 0.3 (mg GAE/g) and 3.53 ± 0.40 (mg QE/g). The % inhibition which was shown by EOs of *R. stricta* was 23.33 ± 0.15 $\mu\text{g}/\text{mL}$. In future characterization of EOs of *R. stricta* can be done by using Raman spectroscopy. And activity of EOs of *R. stricta* such as anticancer and antimicrobial can also be checked.

ACKNOWLEDGEMENT

Authors are thankful to Dr. Abdullah Ijaz Hussain Director Central Hi-Tech Lab, Government College University, Faisalabad for providing facility of GC-FID. Authors are also grateful to Dr. Muhammad Shahid Chairman Biochemistry Department, University of Agriculture, Faisalabad for providing facility of biological activities.

CONFLICTS OF INTEREST

The authors have no conflicts of interest.

REFERENCES

- Abdul-Hameed, Z. H., Alarif, W. M., Sobhi, T. R., Abdel-Lateff, A., Ayyad, S.-E. N., Badria, F. A., & Saber, J. (2021). New cytotoxic indole-type alkaloids obtained from *Rhazya stricta* leaves. *South African Journal of Botany*, 137, 298-302.
- Adams, R. P. (2007). Identification of essential oil components by gas chromatography/mass spectrometry (Vol. 456): Allured publishing corporation Carol Stream, IL.
- Akhgari, A., Laakso, I., Seppänen-Laakso, T., Yrjönen, T., Vuorela, H., Oksman-Caldentey, K.-M., & Rischer, H. (2015). Analysis of indole alkaloids from *Rhazya stricta* hairy roots by ultra-performance liquid chromatography-mass spectrometry. *Molecules*, 20(12), 22621-22634.
- Akyalcin, H., Ozen, F., & Dulger, B. (2006). Anatomy, morphology, palynology and antimicrobial activity of *Amsonia orientalis* Decne. (Apocynaceae) growing in Turkey. *International Journal of Botany*.
- Al-Hasawi, Z., & Al-Harbi, H. (2014). Effect of *Rhazya stricta* dense leaf extract on the liver and kidney tissue structure of albino mice. *Global Advanced Research Journal of Environmental Science and Toxicology*, 3(4), 057-064.
- Anwar, F., Ali, M., Hussain, A. I., & Shahid, M. (2009). Antioxidant and antimicrobial activities of essential oil and extracts of fennel (*Foeniculum vulgare* Mill.) seeds from Pakistan. *Flavour and Fragrance Journal*, 24(4), 170-176.
- Bhadane, B. S., Patil, M. P., Maheshwari, V. L., & Patil, R. H. (2018). Ethnopharmacology, phytochemistry, and biotechnological advances of family Apocynaceae: A review. *Phytotherapy research*, 32(7), 1181-1210.

- Bukhari, N. A., Al-Otaibi, R. A., & Ibbrahim, M. M. (2017). Phytochemical and taxonomic evaluation of *Rhazya stricta* in Saudi Arabia. *Saudi journal of biological sciences*, 24(7), 1513-1521.
- Elkady, A. I. (2013). Crude alkaloid extract of *Rhazya stricta* inhibits cell growth and sensitizes human lung cancer cells to cisplatin through induction of apoptosis. *Genetics and molecular biology*, 36(1), 12-21.
- Elkady, A. I., Hussein, R. A. E. H., & Abu-Zinadah, O. A. J. B. r. i. (2014). Effects of crude extracts from medicinal herbs *Rhazya stricta* and *Zingiber officinale* on growth and proliferation of human brain cancer cell line in vitro. 2014.
- Elkady, A. I. J. G., & biology, m. (2013). Crude alkaloid extract of *Rhazya stricta* inhibits cell growth and sensitizes human lung cancer cells to cisplatin through induction of apoptosis. 36(1), 12-21.
- Elyemni, M., Louaste, B., Nechad, I., Elkamli, T., Bouia, A., Taleb, M., . . . Eloutassi, N. (2019). Extraction of essential oils of *Rosmarinus officinalis* L. by two different methods: Hydrodistillation and microwave assisted hydrodistillation. *The Scientific World Journal*, 2019.
- Hussain, A. I., Anwar, F., Shahid, M., Ashraf, M., & Przybylski, R. (2010). Chemical composition, and antioxidant and antimicrobial activities of essential oil of spearmint (*Mentha spicata* L.) from Pakistan. *Journal of Essential Oil Research*, 22(1), 78-84.
- Hussain, A. I., Anwar, F., Sherazi, S. T. H., & Przybylski, R. (2008). Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food chemistry*, 108(3), 986-995.
- Lanjwani, A. H., Ganghro, A. B., & Khuhawar, T. M. J. (2018). Phytochemical analysis and biological activity of different parts of *Rhazya stricta*. *Rawal Medical Journal*, 43(3), 532-535.
- Lu, J.-J., Bao, J.-L., Chen, X.-P., Huang, M., & Wang, Y.-T. (2012). Alkaloids isolated from natural herbs as the anticancer agents. *Evidence-based complementary and alternative medicine*, 2012.
- Marwat, S. K., Usman, K., Shah, S. S., Anwar, N., & Ullah, I. (2012). A review of phytochemistry, bioactivities and ethno medicinal uses of *Rhazya stricta* Decsne (Apocynaceae). *African Journal of Microbiology Research*, 6(8), 1629-1641.
- Rasool Hassan, B. (2012). Medicinal plants (importance and uses). *Pharmaceut Anal Acta* 3: e139. In.
- Riaz, M., Rasool, N., Bukhari, I. H., Shahid, M., Zahoor, A. F., Gilani, M. A., & Zubair, M. (2012). Antioxidant, antimicrobial and cytotoxicity studies of *Russelia equisetiformis*. *African Journal of Microbiology Research*, 6(27), 5700-5707.
- Silva-Flores, P. G., Pérez-López, L. A., Rivas-Galindo, V. M., Paniagua-Vega, D., Galindo-Rodríguez, S. A., & Álvarez-Román, R. (2019). Simultaneous GC-FID quantification of main components of *Rosmarinus officinalis* L. and *Lavandula dentata* essential oils in polymeric nanocapsules for antioxidant application. *Journal of analytical methods in chemistry*, 2019.
- Sultana, N., & Khalid, A. (2010). Phytochemical and enzyme inhibitory studies on indigenous medicinal plant *Rhazya stricta*. *Natural product research*, 24(4), 305-314.
- Tohidi, B., Rahimmalek, M., & Arzani, A. (2017). Essential oil composition, total phenolic, flavonoid contents, and antioxidant activity of *Thymus* species collected from different regions of Iran. *Food chemistry*, 220, 153-161.