

Seasonal Variation And Diversity Of Fresh Water Microalgae And Cyanobacteria Form Gomuki Dam, Kallakurichi, Tamil Nadu – India

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Abstract

Algae are distributed worldwide in the sea, in freshwater and also in wastewater. Most are microscopic, but some are quite large. In the study the samples were collected from fresh water, moist soils and stone scrapings etc., totally 40 samples were collected in the different seasons from in Gomuki dam located in Villupuram, origin from Kalvarayan hills, Eastern Ghats range Latitude: 11^{*}46'41.21" and Longitude: 78^{*}49'49.88. The samples were screened for the presence of microalgae by plating on BG¹¹, BBM and AC medium. The isolates were further identified based on their morphological characteristics. Among the 40 samples 53 genera and 127 species of various microalgae were identified among that 34 species belong to Bacillariophyceae, 30 species belong to Cyanophyceae, 25 species belong to Chlorophyceae, 12 species belong to Zygnematophyceae, 06 species belong to Conjugatophyceae, 06 species belong to Trebouxiophyceae, 05 species belong to Euglenophyceae, 05 species belong to Mediophyceae, 02 species belong to Coscinodiscophyceae, 01 species belong to Charophyceae, 01 belong to Ulvophyceae families are observed under the microscope. During summer seasons the predominant algal group following that Zygnematophyceae and cyanophyceae was isolated. During rainy seasons the Coscinodiscophyceae were predominant group. During winter most of algae family is less due to climatic changes. The growths are noted under favorable condition for the algal growth. It is the first inclusive taxonomical and diversity study in Gomuki dam and rich of algal species were present also further investigations were needed.

Key words: Algae, Chlorophyta, Bacillariophyta, Cyanoprocaryota, Euglenophyta, Diatoms.

Introduction

Initial researches from 17th century to 18th century the algae were made to define and taxonomic importance. First algae report was a diatom (Tabellaria flocullosa), According to Royal Botanical Society, London, it was collected by an unknown English gentleman in 1703 and found it as epiphytic to roots of an aquatic weed Lemna (Round, 1981). It has been estimated that about 2,00,000 – 8,00,000 species exist of

which about 35,000 species are described by Cheng (2011). There are above 30,000 species of microalgae were isolated and identified as a value-added products that useful in many industrial applications so for by the scientists (Amal, 2021), About 30% work on plant biodiversity and less than 2% of the workforce on microalgae diversity research it is insufficient records and leads to loss of their potential usages (Magurran, 2010).

Algae play an important role in any aquatic ecosystem viz. freshwater bodies, backwaters, saline, estuaries, oceans, effluents and Polar Regions (Narchonai et al., 2019). This organisms is a highly potential, utilized for food, feed, cosmetics, fuel etc., (Nithya et al., 2014). Phytoplanktons not only serve as food, but also play an important role in maintaining the environmental factors and quality of water and contain rich source of protein, lipid and carbohydrate and also highly value added products such as biodiesel, bioethanol, biobutanol, biohydrogen etc (Oscar et al., 2014, Halder et al., 2019).

Freshwater algae of India, there are diverse number of microalgae are reported. The present investigation was carried out as an attempt to assess the diversity status of phytoplankton that present in aquatic ecosystems present in Gomuki dam. It is the first inclusive taxonomical study in dam Gomuki, and rich of algal species are accumulated. Further this study gives a basic idea on utilizing these microalgae for biofuel production and other biotechnological applications.

MATERIALS AND METHODS:

Sampling Site

These areas are seasonal in nature, usually containing water in the months June. These samples are typically distributed along with altitudinal; transects of interest are collected from random positions Gomuki dam. Sample identification was performed according to the morphology of algae.

Collection of samples

40 samples were collected from Gomuki dam seasonal variation (summer, winter, autumn, and rainy seasons), the river origin from Kalvarayan hills. Samples were taken from by scraping rocks, fine gravel, moist soil, pools, riffles, pebbles, dead twigs, roots and stems of some macrophytes. Phytoplankton was collected by towing plankton net into the river. The collected water samples were stored in a sterile 15 ml plastic bottle.

Cultivation of microalgae

The algae samples were scraped and transferred into a sterile plastic bottles using sterile blades and forceps and were transported to the Department of Microbiology, Periyar University, Salem-11, India, for

taxonomic and cultural studies. One set of samples were preserved in 4% formaldehyde and then observed under a light microscope. Samples were transferred to Erlenmeyer flasks containing BG¹¹, BBM medium. They were maintained in culture room.

Isolation of microalgae

The collected each samples were initially inoculated into 100 ml conical flask containing 50ml of BG 11 medium and BBM medium. After inoculation the flasks were incubated at $24+/-2^{\circ}C$ under 37.5 µmol-1m-1sec-1intensity with 16:8h photoperiod for 15 days (Dayananda et al., 2010).

Identification of Microalgae

The isolated microalgae were observed microscopically using light microscope and identified. The fixed algal species were identification in laboratory using compound microscope and texts (Bellinger 2010). The strains were examined under a light microscope. The sample bottle was mixed by shaking, and transferred a volume of 500 μ L for each bottle into a 1.5 ml Eppendorf, a few drops were loaded on a slide and was observed under 400× magnification. Identification of the microalgae species was performed per microscopic fields in the multiple of 10 when no new species have been identified after the 10 fields; the diversity analysis is completed. The identified taxa were subjected to purification using BG ¹¹ medium (Rippka et al., 1979), BBM (Bold, 1973).

Results

The microalgal components of phytoplankton community, 40 samples were collected from Gomuki dam, by scraping rocks, fine gravel, moist soil, pools, riffles, pebbles, dead twigs, roots and stems of some macrophytes, were diverse. Taxonomic evaluation of the collected material indicated the presence of 53 genera (sample collection site - Fig 1).



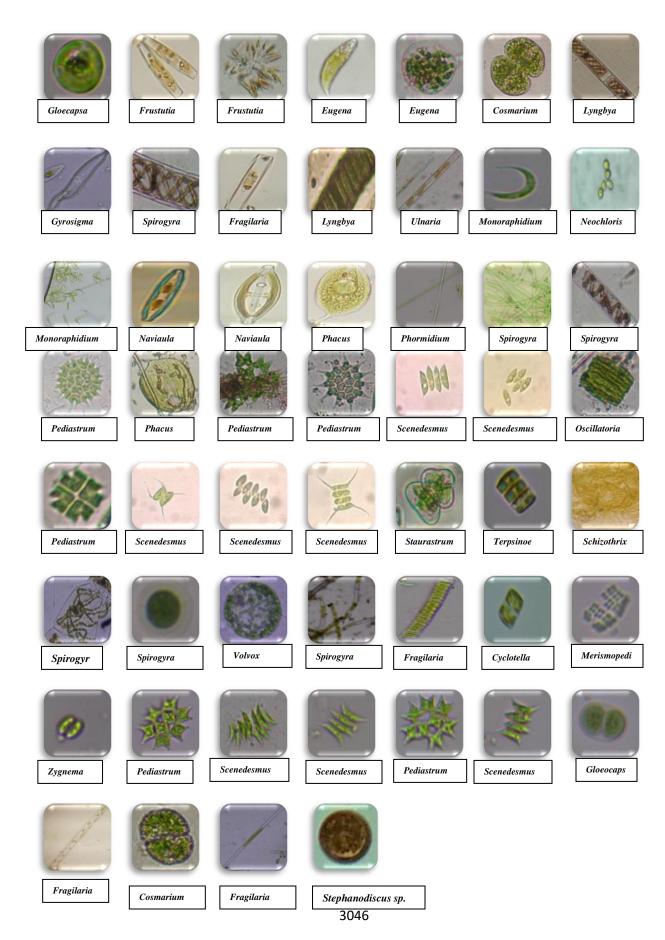
Fig. 1: Site of the study area

Among the 40 samples, 53 Genera and 127 species of various microalgae were identified and observed under microscope (Fig 2). In this study showed that the temperature ranges from 28-36°C (in water 25-34°C); pH ranges from 6-7.5 were recorded (Table 1).

SL	Parameters	Water
1	Colour	Almost colorless
2	Odour	Almost no smell
3	Atmospheric temperature	20°C to 37°C
4	Water temperature	15.1°C to 30.5°C
5	Turbidity	Clear
6	рН	6.8
7.	Nature	Drinking & irrigation

Table-1: The parameters of selected were discussed in the localities of Gomuki dam, Kallakurichi (DT), India.







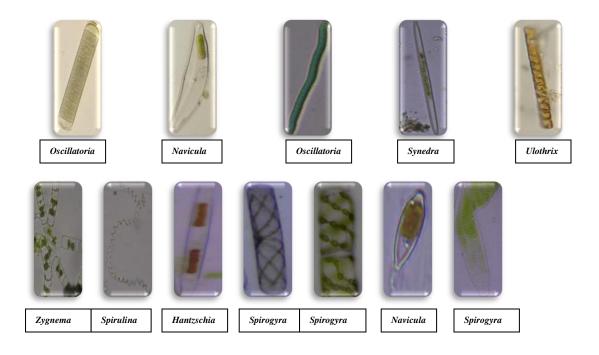


Fig. 2: Microscopic image of microalgae

Among that 127 species belong to 11 families of 34 (26.77%) isolates belong to the family Bacillariophyceae, 30 (23.62%) belong to the family Cyanophyceae, 25 (19.68%) belong to Chlorophyceae, 12 (09.44%) belong to the family of Zygnematophyceae, 06 (04.72%) belong to Conjugatophyceae, 06 (04.72%) belong to Trebouxiophyceae, 05 (03.93%) belong to Mediophyceae families, 5 (03.93%) belong to Euglenophyceae, 02 (01.57%) belong to Coscinodiscophyceae families, 1 (0.78%) belong to Charophyceae families are observed under the light microscope. The algae are described below (Fig:3, Table 2&3).

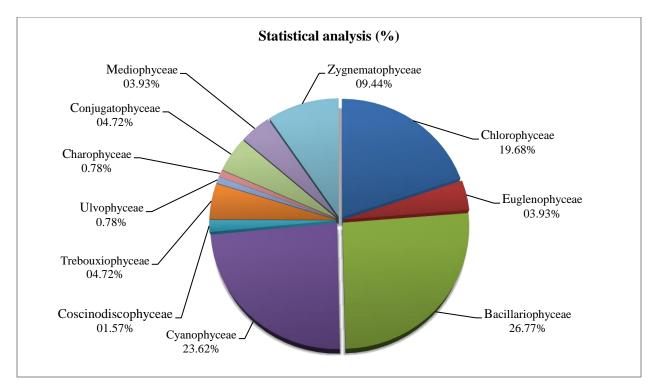


Fig 3. Algae diversity of study area

SL	Class	Number of	Statistical	Number of	Statistical
		Genera	analysis (%)	Species	analysis (%)
1	Chlorophyceae	10	18.86	25	19.68
2	Euglenophyceae	02	03.77	05	03.93
3	Bacillariophyceae	13	24.52	34	26.77
4	Cyanophyceae	13	24.52	30	23.62
5	Coscinodiscophyceae	02	03.77	02	01.57
6	Trebouxiophyceae	03	05.66	06	04.72
7	Ulvophyceae	01	01.88	01	00.78
8	Charophyceae	01	01.88	01	00.78
9	Conjugatophyceae	03	05.66	06	04.72
10	Mediophyceae	03	05.66	05	03.93
11	Zygnematophyceae	02	03.77	12	09.44
	Total	53	99.87	127	99.96

Table-2: Statistical analysis of genera and species of various algal classes.

S. No	Class		Genus	No of species
		1.	Hantzschia	2
		2.	Nitzschia	2
		3.	Cymbella	3
		4.	Gomphonema	1
		5.	Fragilaria	7
		6.	Synedra	1
1	Bacillariophyceae	7.	Ulnaria	2
		8.	Gyrosigma	2
		9.	Navicula	5
		10.	Frustulia	2
		11.	Pleurosigma	3
		12.	Pinnularia	3
		13.	Meridion	1
2	Coscinodiscophyceae	14.	Coscinodiscus	1
		15.	Melosira	1
		16.	Terpsinoe	1
3	Mediophyceae	17.	Cyclotella	3
		18.	Stephanodiscus sp.	1
4	Charophyceae	19.	Chara	1
		20.	Cosmarium	4
5	Conjugatophyceae	21.	Staurastrum	1
		22.	Closterium	1
		23.	Coelastrum	1
		24.	Scenedesmus	8
		25.	Characium	1
		26.	Pediastrum	6
		27.	Neochloris	1
6	Chlorophyceae	28.	Monoraphidium	2
		29.	Chlorococcum	3

		30.	Characiochloris	1
		31.	Volvox	1
		32.	Haematococcus	1
		33.	Dictyosphaerium	1
7	Trebouxiophyceae	34.	Chlorella	3
		35.	Oocystis	2
8	Ulvophyceae	36.	Ulothrix	1
		37.	Chroococcus	3
		38.	Gloeocapsa	2
		39.	Oscillatoria	6
		40.	Phormidium	6
9	Cyanophyceae	41.	Lyngbya	3
		42.	Dermorcarpa	1
		43.	Schizothrix	1
		44.	Aphanocapsa	1
		45.	Merismopedi	2
		46.	Nostoc	1
		47.	Fremyella	1
		48.	Calothrix	2
		49.	Spirulina	1
10	Euglenophyceae	50.	Euglena	3
		51.	Phacus	2
11	Zygnematophyceae	52.	Zygnema	2
		53.	Spirogyra	10
	Total	·		127

 Table-3: Number of genera and species.

S.	Phylum	Class	Order	Family	Genus
No					
			Bacillariales	Bacillariaceae	Hantzschia,

					Nitzschia
				Cymbellaceae	Cymbella
			Cymbellales	Gomphonematace	Gomphonema
				ае	
			Fragilariales	Fragilariaceae	Fragilaria, Synedra
		Bacillariophyceae	Licmophorales	Ulnariaceae	Ulnaria
				Naviculaceae	Gyrosigma,
					Navicula
1	Bacillariophyta		Naviculales	Amphipleuraceae	Frustulia
				Pleurosigmatacea	Pleurosigma
				е	
				Pinnulariaceae	Pinnularia
			Tabellariales	Tabellariaceae	Meridion
		Coscinodiscophyc	Coscino-	Coscinodiscaceae	Coscinodiscus
		eae	discales		
			Melosirales	Melosiraceae	Melosira
			Anaulales	Anaulaceae	Terpsinoe
		Mediophyceae	Stephano-	Stephanodiscacea	Cyclotella,
			discales	e	Stephanodiscus
		Charophyceae	Charales	Characeae	Chara
				Desmidiaceae	Cosmarium,
2	Charophyta		Desmidiales		Staurastrum
		Conjugatophyceae		Closteriaceae	Closterium
			Zygnematales	Zygnemataceae	Spirogyra, Zygnema
				Scenedesmaceae	Scenedesmus,
					Coelastrum
			Sphaeropleales	Characiaceae	Characium
		Chlorophyceae		Hydrodictyaceae	Pediastrum
				Neochloridaceae	Neochloris
				Selenastraceae	Monoraphidium
			Chlamydo-	Chlorococcaceae	Chlorococcum

3	Chlorophyta		monadales	Characio-	Characiochloris
				chloridaceae	
				Chlorellaceae	Dictyochloropsis,
					Chlorella,
		Trebouxiophyceae	Chlorellales	Haemato-	Haematococcus
				coccaceae	
				Oocystaceae	Oocystis
		Ulvophyceae	Ulotrichales	Ulotrichaceae	Ulothrix, Volvox
			Chroococcales	Chroococcaceae	Chroococcus
				Microcystaceae	Gloeocapsa
			Oscillatoriales		Oscillatoria,
				Oscillatoriaceae	Phormidium,
					Lyngbya, Spirulina
			Pleurocapsales	Dermocarpellacea	Dermorcarpa
4	Cyanobacteria	Cyanophyceae		e	
				Synechococcales	Schizothrix
			Synecho-	Merismopediacea	Aphanocapsa,
			coccales	e	Merismopedi
				Nostocaceae	Nostoc
			Nostocales	Microchaetaceae	Fremyella
				Rivulariaceae	Calothrix
5	Euglenoidea	Mastigophyceae	Euglenoidi-	Euglenoidaceae	Euglena
			dales		
6	Euglenozoa	Euglenophyceae	Euglenales	Phacaceae	Phacus

Table: 4 Classification of algae group

Based on the seasonal variation the microalgae were listed. For the growth of algae the pH and temperature is most important factor. Due to the climatic changes the algae grown rates are variable (Table 5).

SI. No.	Seasons pH		Temperature °C
	(Four)	(Acidic, Neutral, Acid)	(0 to 100 °C)

1	Summer	6.7 - 6.9	29 - 35°C
	(Mar, Apr, May		
2	Raining	6.5 - 6.8	25 - 29°C
	(Jun, Jul, Aug)		
3	Autumn	7.0 - 7.5	20 - 25°C
	(Sep, Oct, Nov)		
4	Winter	7.7 - 8.0	15 - 20°C
	(Dec, Jan, Feb)		

Table-5: The parameters of pH and Temperature during four seasons

Most of the bacillariophyceae family was grown during summer following this cyanophyceae, Zygnematophyceae, Trebouxiophyceae, chlorophyceae, Mediophyceae. During raining seasons the Coscinodiscophyceae were predominant growth. Cyanophyceae were grown mostly in autumn and summer time. During winter the algae growth rate was less due to climatic changes (Table 6).

SI.	Family	Summer	Raining	Autumn	Winter
No.		(Mar, Apr, May)	(Jun, Jul, Aug)	(Sep, Oct, Nov)	(Dec, Jan, Feb)
1	Bacillariophyceae	*****	xxx	×	××
2	Cyanophyceae	ддддд	дд	адада	¤
3	Chlorophyceae	***	**	**	*
4	Zygnematophyceae	33333	Э	-	33
5	Conjugatophyceae	000	-	۵۵	-
6	Trebouxiophyceae	¥¥¥¥	¥¥¥	¥¥	¥
7	Euglenophyceae	≈≈≈	≈≈≈	~	-
8	Mediophyceae	0000	$\diamond \diamond$	000	-
9	Coscinodiscophyceae	0	0000	-	00
10	Ulvophyceae	•••	•	-	•
11	Charophyceae	##	-	-	#

Table-6: Seasonal variation of Microalgal diversity

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The algae isolated from fresh water are Aphanocapsa sp., Calothrix sp., Cosmarium sp., Hantzschia sp., Nostoc sp., Phormidium sp., Schizothrix sp., Characiochloris sp., Chrococcus sp., Cymbella sp., Dictyochloropsis sp., Euglena sp., Oscillatoria sp., Pinnularia sp., Chlorococcum sp., Chlorella sp., Coelastrum sp., Oocystis sp., Scenedesmus sp., Closterium sp., Fragilaria sp., Melosira sp., Nitzschia sp., Spirogyra sp., Characium sp., Coscinodicscus sp., Cyclotella sp., Navicula sp., Chara sp., Dermorcarpa sp., Terpsinoe sp., Fremyella sp., Frustulia sp., Gloeocapsa sp., Gyrosigma sp., Gomphonema sp., Lyngbya sp., Phacus sp., Pleurosigma sp., Spirulina sp., Synedra sp., Haematococcus sp., Meridion sp., Monoraphidium sp., Merismopedia sp., Staurastrum sp., Pedistrum sp., Neochloris sp., Stephanodiscus sp., Ulothrix sp., Ulnaria sp., Volvox and Zygnema sp.,

Discussion

This is the first report the algae diversity in the Kalvarayan hills of Gomuki dam. There are also only few papers dealing with epilithic micoralgae of high mountain lakes and rock moist soil from other regions in Tamilnadu hill station (Suresh et al., 2012). The Western Ghats is one of the hotspot of biodiversity (Myers et al., 2000). Narmada river contain mostly of Chlorophyceae, Bacillariophyceae, Cyanophyceae, Euglenophyceae, the population of plankton vary in different seasons and months Sohani (2015).

In our study from 53 genera 127 species of microalgae were identified in this study. The diversity index showed that Bacillariophyceae members were dominant followed by Cyanophyceae, Chlorophyceae, Zygnematophyceae, Conjugatophyceae, Trebouxiophyceae Euglenophyceae Mediophyceae Coscinodiscophyceae Ulvophyceae and Charophyceae. In previous study a total of 39 genera of 70 species of microalgae diversity index showed that Chlorophyceae members were dominant followed by Cyanophyceae and Bacillariophyceae (Narchonai et al., 2021).

Cyanophyceae 4, Chlorophyceae 3 Bacillariophyceae 2, Trebouxiophyceae 2, Conjugatophyceae 1 and Euglenophyceae 1 results obtained from previous study of Amal, (2021).

Similar result was obtained earlier in another study Chlorophyceae and Cyanophycean genera are more abundant in winter season than in late summer (Halder et al., 2019). Bacillariophyceae Navicula sp., Pinnularia sp., Euglenophyceae Euglena sp., Phacus sp., (Ajayan, 2015).

Jindal et al., 2014, studied the diversity of Kuntbhyog lake total 48 species of planktons were repoted. Bacillariophyceae 55.98% and Chlorophyceae 27.68% formed the dominant variety, and following Cyanophyceae 13.49% and Euglenophyceae 2.85%.

Thakur et al., 2013, from three freshwater lakes 9 groups of algae having 148 species were studied over period of two years. Jha et al., 1985, had documented 65 Chloropytes and one Euglenophyte from

Govindsagar reservoir. From Jharkhand, India, 25 taxa of cyanobaacterial species rich in fresh water habitat of Bokaro thermal power station. It has been reported for the first time from this area (Arpana 2010).

44 algal species were recorded, chlorophyceae, bacillariophyceae, cyanophyceae. Since, the reservoir shows the presence of various algal species, it should be protected as a natural wealth (Ashish 2015). In total 35 chlorophytes, 10 cyanophytes and 4 bacillariophytes were isolated and described (Mohanapriya 2014).

42 species of Chlorococcales were recorded and described in area during January 2014 to Dec. 2015 reported in Lakhpat. In three sites total 25 taxa of class Cyanophyceae were recorded for the first time from Yelderi dam (Mulani 2015). There were 28 species of blue-green algae that belong to Mougeotia species and Spirogyra observed in Kuttanad, (Smitha 2017).

Water samples collected from Tiruchirappalli India, the six chosen sites during March 2014, 81 species of 35 species belongs to Chlorophyta, 9 species belongs to Bacillariophyta and 37 species belongs to Cyanophyta. 8 microalgal genera found predominant in the lake (Vijayan et al., 2014).

During summer month of June, 2004 sampling of 5 different freshwater ponds in and around from Thanjavur, Tamilnadu. Physico-chemical parameters of pond waters were analyzed and totally 39 species of cyanobacteria were recorded (Muthukumar et al., 2007). Chlorococcales reported from different region of India by many researches like, Bharat (1964); Still now so many workers did the study on algal biodiversity (Ambika, 2016).

Conclusion

Present study shows the richness microalgal species based on temperature, light intensity and quality of fresh water based on the seasonal variation. Variation of cellular organization prokaryotes to eukaryotes was noticed. In morphology there is great organization diversity from unicellular, colonial to coenobium form unbranched to branched thallus. Monitoring the algae diversity in the river provides the water quality, environmental pollution and the impact of human activity on natural resources, and enable conservation biology. Thus in a water body how this mechanic diversity occur is a matter of further study.

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References

1 Ajayan AP., KG AK. (2015). Micro algal diversity of the lake inside the government zoological garden, Thiruvanathapuram, Kerala India. Inter. J of Envi Sci; 6 (3): 330- 337.

2 Round F.E. (1981). The ecology of algae. New York: Cambridge University press.

3 Cheng KC., Ogden KI. (2011). Algal biofuels the research, American institute of chemical engineers (aiche).

4 Amal AB., Fouad WM. (2021). Identification of culturable microalgae diversity in the River Nile in Egypt using enrichment media, Afr J Bio Sc, 3(2): 50-64.

5 Magurran, AE., Dornelas, M. (2010). Biological diversity in a changing world. Philosophical Transactions of the Royal Society B: Biological Sciences, 365(1558), 3593-3597.

6 Narchonai G., Arutselvan C., Lewis OF., Thajuddin N. (2019). Deciphering the microalgal diversity and water quality assessment of two urban temple ponds in Pondicherry, India Biocatalysis and Agricultural Biotechnology, 22: 1-9, 101427.

7 Nithya C., Lewisoscar F., Kanaga S., Kavitha R., Bakkiyaraj D., Arunkumar M., Naiyf Alharbi S., Chinnathambi A., Alharbi AS., Thajuddin N. (2014). Biofilm inhibitory potential of Chlamydomonas sp. Extract against P. aeruginosa. J Algal biomass utln. 5 (4): 74-81.

8 Oscar FL., Bakkiyaraj D., Nithya C., Thajuddin N. (2014). Deciphering the diversity of microalgal bloom in wastewater-an attempt to construct Potential consortia for bioremediation. J. Current Perspectives in Appl. Microbio. 2278, 92.

9 Dayananda C., Kumudha A., Sarada R., Ravishankar GA. (2010). Isolation, characterization & outdoor cultivation of green microalgae Botryococcus sp. ERS, 5: 2497-2505.

10 Bellinger EG., Sigee DC. (2010). Freshwater Algae: Identification and use as bioindicators. In Freshwater Algae: Identification and Use as Bioindicators. Wiley-Blackwell.

11 Rippka RJ., Deruelles JB., Waterbury M., Herdman RY., Stanier. (1979). Generic assignments, strain histories and properties of pure cultures of BGA. Microbiology, 111: 1-61.

12 Bold HC. (1973). Morphology of plants. 3rd edition. New York: Harper and Row.

13 Suresh A., Preeven Kumar R., Dhanasekaran D., Thajuddin N. (2012). Biodiversity of microalgae Western and Eastern ghats, India. Pakistan j of biological j, 15 (19): 919-928.

14 Sohani S. (2015). Diversity of fresh water algae in river narmada at jalud (mandleshwer) Indore, India research journal of recent sciences, 4: 14-17.

15 Smitha S. (2017). Filamentous algal diversity in Kole wetland paddy fields. Int J Adv Res, 5(6): 361-366.

16 Thakur RK., Jindal R., Singh UB., Ahluwalia AS. (2013). Plankton diversity and water quality assessment of three freshwater lakes of Mandi (India) with special reference to planktonic indicators. Environ Monit Assess, 185(10): 8355-8373.

17 Vijayan D., Manivannan K., Santhoshkumar S., Pandiaraj D., Mohamedimran M., Thajuddin N., Kala K., Muhammad MH. (2014). Depiction of microalgal diversity in Gundur lake, Tiruchirappalli (Dt) Tamil Nadu, India. Asian j of biolo sci, 7: 111-121.

18 Arpana S. (2010). Cyanobacterial flora from fresh water habitat of Bokaro thermal power station (dist. Bokaro) Jharkhand, India. Int J of plant sciences, 5(2): 679-681.

19 Ashish W., Yenkar (2015). Bio-diversity of fresh water algae of Rotha-ii reservoir of Wardha district of Maharashtra, India. IJRSB, 3(11): 30-32.

20 Ambika HD., Krishnamurthy SR. (2016). Diversity and species composition of subaerial algae in Kuvempu University campus, Shimoga, Karnataka, India. The JIBS, 95: 78-91.

21 Bharati S.G. (1964). Chlorococcales from Kodai Kanal, South India. J Bombay Nat Hist Soc, 61: 475-479.

22 Jha B.C, Kaushal D.K., Rama R. (1985). Chlorococeales of Govindsagar reserviour, HP. India Phykos, 24: (1& 2): 27-32.

23 Jindal R., Thakur RK., Uday BS., Ahluwalia AS. (2014). Phytoplankton dynamics & water quality of Prashar Lake, HP, India sustainabil. of water quality and ecology, 3(4): 101-113.

24 Muthukumar C., Muralitharan G., Vijayakumar R., Panneerselvam A., Thajuddin N. (2007). Cyanobacterial biodiversity from different freshwater ponds of Thanjavur, Tamil Nadu (India). Acta botanica malacitana, 32: 17-25.

25 Mohanapriya K.R., Geetharamani D. (2014). Fresh water microalgal diversity of Noyyal river at Tamil Nadu State, India. J Algal Biomass Utln, 5(4): 12-20.

26 Myers N., Mittermeier R.A., Mittermeier C.G., Fonseca G.A.B., Kent J. (2000). Biodiversity hotspots for conservation priorities. Nature, 403: 853-858.

27 Halder P., Debnath M., Ray S. (2019). Occurrence and diversity of microalgae in phytoplankton collected from freshwater community ponds of Hooghly District, West Bengal, India. Plant Science Today, 6(1): 8-16.

28 Mulani R.M., Sonule M.D. (2015). Fresh water cyanophycean algae from Velderi dam Parbhani district (MS.), India. International j of science and research, 4(1): 740-742.